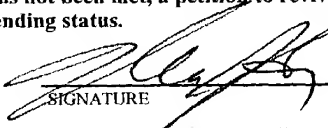


30 APR 2001

FORM PTO-1390 (REV. 11-2009)		U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE		ATTORNEY'S DOCKET NUMBER 960296.96617	
TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371				U.S. APPLICATION NO. (If known, see 37 CFR 1.5)	
				09/830751	
INTERNATIONAL APPLICATION NO. PCT/US00/23878		INTERNATIONAL FILING DATE 30 August 2000 (30.08.00)		PRIORITY DATE CLAIMED 30 August 1999 (30.08.99)	
TITLE OF INVENTION Production of 3-Hydroxypropionic Acid in Recombinant Organisms					
APPLICANT(S) FOR DO/EO/US SUTHERS, Patrick F. and CAMERON, Douglas C.					
Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:					
<p>1. <input checked="" type="checkbox"/> This is a FIRST submission of items concerning a filing under 35 U.S.C. 371.</p> <p>2. <input type="checkbox"/> This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371.</p> <p>3. <input type="checkbox"/> This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (21) indicated below.</p> <p>4. <input type="checkbox"/> The US has been elected by the expiration of 19 months from the priority date (Article 31).</p> <p>5. <input checked="" type="checkbox"/> A copy of the International Application as filed (35 U.S.C. 371(c)(2))</p> <p style="margin-left: 20px;">a. <input type="checkbox"/> is attached hereto (required only if not communicated by the International Bureau).</p> <p style="margin-left: 20px;">b. <input type="checkbox"/> has been communicated by the International Bureau.</p> <p style="margin-left: 20px;">c. <input checked="" type="checkbox"/> is not required, as the application was filed in the United States Receiving Office (RO/US).</p> <p>6. <input type="checkbox"/> An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).</p> <p style="margin-left: 20px;">a. <input type="checkbox"/> is attached hereto.</p> <p style="margin-left: 20px;">b. <input type="checkbox"/> has been previously submitted under 35 U.S.C. 154(d)(4).</p> <p>7. <input checked="" type="checkbox"/> Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))</p> <p style="margin-left: 20px;">a. <input type="checkbox"/> are attached hereto (required only if not communicated by the International Bureau).</p> <p style="margin-left: 20px;">b. <input type="checkbox"/> have been communicated by the International Bureau.</p> <p style="margin-left: 20px;">c. <input type="checkbox"/> have not been made; however, the time limit for making such amendments has NOT expired.</p> <p style="margin-left: 20px;">d. <input checked="" type="checkbox"/> have not been made and will not be made.</p> <p>8. <input type="checkbox"/> An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371 (c)(3)).</p> <p>9. <input type="checkbox"/> An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).</p> <p>10. <input type="checkbox"/> An English language translation of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).</p> <p>Items 11 to 20 below concern document(s) or information included:</p> <p>11. <input type="checkbox"/> An Information Disclosure Statement under 37 CFR 1.97 and 1.98.</p> <p>12. <input type="checkbox"/> An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.</p> <p>13. <input type="checkbox"/> A FIRST preliminary amendment.</p> <p>14. <input type="checkbox"/> A SECOND or SUBSEQUENT preliminary amendment.</p> <p>15. <input type="checkbox"/> A substitute specification.</p> <p>16. <input type="checkbox"/> A change of power of attorney and/or address letter.</p> <p>17. <input type="checkbox"/> A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821 - 1.825.</p> <p>18. <input type="checkbox"/> A second copy of the published international application under 35 U.S.C. 154(d)(4).</p> <p>19. <input type="checkbox"/> A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).</p> <p>20. <input checked="" type="checkbox"/> Other items or information:</p> <p style="margin-left: 40px;">Postcard receipt</p>					

JC18 Rec'd PCT/PTO 3 0 APR 2001

U.S. APPLICATION NO. (37 CFR 1.53) 097/830751		INTERNATIONAL APPLICATION NO. PCT/US00/23878		ATTORNEY'S DOCKET NUMBER 960296.96617	
21. <input checked="" type="checkbox"/> The following fees are submitted: BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)): Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO. \$1000.00 International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$860.00 International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$710.00 International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$690.00 International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) \$100.00 ENTER APPROPRIATE BASIC FEE AMOUNT =				CALCULATIONS PTO USE ONLY <div style="border: 1px solid black; padding: 2px;">\$ 710</div>	
Surcharge of \$130.00 for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(e)).				<div style="border: 1px solid black; padding: 2px;">\$</div>	
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE	\$	
Total claims	- 20 =		x \$18.00	\$	
Independent claims	6 - 3 =	3	x \$80.00	\$ 240	
MULTIPLE DEPENDENT CLAIM(S) (if applicable)				+ \$270.00	
TOTAL OF ABOVE CALCULATIONS =				\$ 950	
<input type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2.				+ \$ 0	
SUBTOTAL =				\$ 950	
Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f)).				<div style="border: 1px solid black; padding: 2px;">\$ 0</div>	
TOTAL NATIONAL FEE =				\$ 950	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property +				<div style="border: 1px solid black; padding: 2px;">\$ 0</div>	
TOTAL FEES ENCLOSED =				\$ 950	
				Amount to be refunded:	\$
				charged:	\$
a. <input type="checkbox"/> A check in the amount of \$ _____ to cover the above fees is enclosed. b. <input checked="" type="checkbox"/> Please charge my Deposit Account No. <u>17-0055</u> in the amount of \$ <u>950</u> to cover the above fees. A duplicate copy of this sheet is enclosed. c. <input checked="" type="checkbox"/> The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. <u>17-0055</u> . A duplicate copy of this sheet is enclosed. d. <input type="checkbox"/> Fees are to be charged to a credit card. WARNING: Information on this form may become public. Credit card information should not be included on this form. Provide credit card information and authorization on PTO-2038.					
NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137 (a) or (b)) must be filed and granted to restore the application to pending status.					
SEND ALL CORRESPONDENCE TO:					
Nicholas J. Seay QUARLES & Brady LLP P.O. Box 2113 Madison, WI 53701-2113			<div style="text-align: center;">  SIGNATURE _____ Nicholas J. Seay NAME _____ 27,386 REGISTRATION NUMBER </div>		

PRODUCTION OF 3-HYDROXYPROPIONIC ACID IN RECOMBINANT
ORGANISMS

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims priority from U.S. Provisional Patent Application S.N.

5 60/151,440 filed August 30, 1999.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH
OR DEVELOPMENT

The research project which gave rise to the invention described in this patent application was supported by EPA grant R824726-01. The United States Government
10 may have certain rights in this invention.

BACKGROUND OF THE INVENTION

The technology of genetic engineering allows the transfer of genetic traits between species and permits, in particular, the transfer of enzymes from one species to others. These techniques have first reached commercialization in connection with high-
15 value added products such as pharmaceuticals. The techniques of genetic engineering are equally applicable and cost effective when applied to genes and enzymes which can be used to make basic chemical feedstocks.

A metabolic pathway of interest exists in the bacteria *Klebsiella pneumoniae*, which has the ability to biologically produce 3 - hydroxypropionaldehyde from glycerol.
20 Native microorganisms have the ability to produce 1,3 - propanediol from glycerol as well. Commercial interests are exploring the production of 1,3 - propanediol from glycerol or glucose, in recombinant organisms which have been engineered to express the enzymes necessary for 1,3 - propanediol production from other organisms.

3 - hydroxypropionic acid CAS registry Number [503-66-2] (abbreviated as 3-
25 HP) is a three carbon non-chiral organic molecule. The IUPAC nomenclature name for

this molecule is propionic acid 3 - hydroxy. It is also known as 3 - hydroxypropionate, β - hydroxypropionic acid, β - hydroxypropionate, 3 - hydroxypropionic acid, 3 - hydroxypropanoate, hydracrylic acid, ethylene lactic acid, β -lactic acid and 2 - deoxyglyceric acid. Applications of 3-HP include the manufacture of absorbable
5 prosthetic devices and surgical sutures, incorporation into beta-lactams, production of acrylic acid, formation of trifluoromethylated alcohols or diols, polyhydroxyalkonates, and co-polymers with lactic acid. 3-HP for commercial use is now commonly produced by organic chemical syntheses. The 3-HP produced and sold by these methods is relatively expensive, and it would be cost prohibitive to use it for the production of
10 monomers for polymer production. As discussed below, some organisms are known to produce 3-HP. However, there is not yet available a catalog of genes from these organisms and thus the ability to synthesize 3-HP using the enzymes natively responsible for the synthesis of that molecule in the native hosts which produce it does not now exist.

15 In addition to its commercial utility, 3-HP it is found in a number of biological processes, notably including many naturally occurring bio-polymers. Poly(3 - hydroxybutyrate) (PHB) is the most abundant member of the microbial polyesters which contain hydroxy monomers termed polyhydroxyalkonates (PHAs). PHB has utility as a biodegradable thermoplastic material and the material was first produced industrially in
20 1982.

The majority of published research on PHA's that contain 3-HP has concentrated on two bacterial sources: *Ralstonia eutropha* ("*Alcaligenes eutrophus*") and *Pseudomonas oleovorans*. Both *Ralstonia eutropha* and *Pseudomonas oleovorans* are able to grow on a nitrogen free media containing 3 - hydroxy - propionic acid, 1,5 -
25 pentanediol or 1,7 - heptanediol. When 3-HP is the major hydroxy-acid added to the growth media, poly(3 - hydroxybutyrate - co - 3 - hydroxypropionic acid) is formed containing 7 mol % 3 - hydroxypropionic acid. These cells also store 3 mol %, 3 - hydroxypropionic acid poly(3 - butyrate - co - 3 - hydroxypropionic acid).

Recombinant systems have been used to create PHAs. An *E. coli* strain
30 engineered to express PHA synthase from either *Ralstonia eutropha* or *Zoogoea ramigera* produced poly(3 - hydroxypropionic acid) when feed 1,3 - propanediol.

Skraly, F. A. "Polyhydroxyalkanoates Produced by Recombinant *E. coli*." Poster at Engineering Foundation Conference: Metabolic Engineering II, 1998. An *E. coli* strain that expressed PHA synthase (MBX820), when provided with the genes encoding glycerol dehydratase and 1,3 - propanediol dehydratase from *K. pneumonia*, and 4 -
5 hydroxybutyral- CoA transferase from *Clostridium kluyveri*, synthesized PHB from glucose.

Glycerol dehydratase, found in the bacterial pathway for the conversion of glycerol to 1,3 - propanediol, catalyzes the conversion of glycerol to 3 - hydroxypropionaldehyde and water. This enzyme has been found in a number of
10 bacteria including strains of *Citrobacter*, *Klebsiella*, *Lactobacillus*, *Enterobacter* and *Clostridium*. In the 1,3 - propanediol pathway a second enzyme 1,3 - propanediol oxido-reductase (EC 1.1.202) reduces 3 - hydroxypropanaldehyde to 1,3 - propanediol in a NADH dependant reaction. The pathway for the conversion of glycerol to 1,3 - propanediol has been expressed in *E. coli*. Tong et al., Applied and
15 Environmental Microbiology 57 (12) 3541-3546. The genes responsible for the production of 1,3 - propanediol were cloned from the *dha* regulon of *Klebsiella pneumoniae*. Glycerol is transported into the cell by the glycerol facilitator, and then converted into 3 - hydroxy - propionaldehyde by a coenzyme B₁₂- dependent dehydratase. *E. coli* lacks a native *dha* regulon, consequently *E. coli* cannot grow
20 anaerobically on glycerol without an exogenous electron acceptor such as nitrate or fumarate.

Aldehyde dehydrogenases are enzymes that catalyze the oxidation of aldehydes to carboxylic acids. The genes encoding non-specific aldehyde dehydrogenases have been identified in a wide variety of organisms e.g.; *ALDH2* from *Homo sapiens*, *ALD4*
25 from *Saccharomyces cerevisiae*, and from *E. coli* both *aldA* and *aldB*, to name a few. These enzymes are classified by co-factor usage, most require either AND⁺, or NADP⁺ and some will use either co-factor. The genes singled out for mention here are able to act on a number of different aldehydes and it likely that they may be able to oxidize 3 - hydroxy - propionaldehyde to 3 - hydroxypropionic acid.

BRIEF SUMMARY OF THE INVENTION

The present invention is intended to permit the creation of a recombinant microbial host which is capable of synthesizing 3-HP from a starting material of glycerol or glucose. The glycerol or glucose is converted to 3 -

5 hydroxypropionaldehyde (abbreviated as 3-HPA) which is then converted to 3-HP.

This process requires the so-called *dhaB* gene from *Klebsiella pneumoniae* which encodes the enzyme glycerol dehydratase any one of four different aldehyde dehydrogenase genes to convert 3-HPA to 3-HP. The four aldehyde dehydrogenase genes used were *aldA* from the bacterium *E. coli*, *ALDH2* from humans, *ALD4* from the
10 yeast *Saccharomyces cerevisiae*, and *aldB* from *E. coli*. The yeast gene appeared to give the best results.

It is an object of the present invention to provide a genetic construct which encodes glycerol dehydratase and aldehyde dehydrogenase enzymes necessary for the production of 3 - hydroxypropionic acid from glycerol.

15 It is also an object of the present invention to provide a method for the production of 3 - hydroxypropionic acid from glycerol.

Other features and advantages of the invention will be apparent from the following description of the preferred embodiment thereof and from the claims.

20 BRIEF DESCRIPTION OF THE SEVERAL VIEWS OF THE DRAWINGS

Not applicable.

DETAILED DESCRIPTION OF THE INVENTION

It is disclosed here that it is possible to introduce into a bacterial host genes encoding two enzymes and thus confer upon that host the ability to produce 3-HP from
25 glycerol. The two necessary enzymes are glycerol dehydratase and aldehyde dehydrogenase. It is here reported that the two enzymes are both necessary and sufficient to enable a strain of a suitable host, such as a competent *E. coli* strain, to make 3-HP from glycerol. An exemplary gene encoding a glycerol dehydratase is known, the *dhaB* gene from *Klebsiella pneumoniae*, sequenced and rendered convenient to use.
30 Several exemplary aldehyde dehydrogenases are known, and their sequences are

presented here. From this information, it becomes practical to confer upon a bacterial host the ability to convert glycerol into 3-HP in a commercially reasonable manner.

It was not apparent before the completion of the work described here that these two diverse enzymes could be produced in a common host to produce the ability to
5 make 3-HP. There are many known aldehyde dehydrogenase enzymes and genes, and the enzymes are known to have varying substrate specificities and efficiencies. There was not evidence, prior to the work described here, that the aldehyde dehydrogenase enzyme would work on the 3-hydroxypropionaldehyde (3-HPA) substrate to create 3-HP. Without that knowledge, there was no data from which to predict the effectiveness
10 of the 3-HP production studies described below. An additional uncertainty arises from the fact that the intermediate aldehyde, 3-HPA, is toxic to many bacterial host and thus the survival of the host is dependent upon the relative rates of enzymatic production and conversion of the aldehyde intermediate to non-toxic 3-HP.

A difficulty in the realization of the production of 3-HP desired here is that
15 ribosome binding sites from non-native hosts are often ineffectual and lead to poor protein production and that many non-native promoters are often poorly transcribed and a bar to high protein expression. However, the inventors also recognized that a non-native promoter that is known to be very active and is inducible by the addition of a small molecule unrelated to the pathway being expressed is often a very efficient way to
20 express and regulate the levels of enzymes expressed in hosts such as *E. coli*. To achieve high levels of regulated gene expression plasmids were constructed which placed the expression of all exogenous genes necessary for the production of 3 - hydroxypropionic acid from glycerol under the regulation of the *trc* promoter. The *trc* promoter, is efficient, not native to *E. coli*, and inducible by the addition of IPTG.

25 The present specification describes a genetic construct for use in the production of 3 - hydroxypropionic acid from glycerol. The genetic construct includes exemplary DNA sequences coding for the expression of a glycerol dehydratase and a DNA sequence coding for aldehyde dehydrogenase. The set of exemplary sequences necessary for the expression of glycerol dehydratase is collectively referred to as
30 "*dhaB*". The set of sequences necessary for the expression of aldehyde dehydrogenase includes any one of four different genes which proved efficacious. The individual

aldehyde dehydrogenase sequences referred to individually as *ALDH4*, *ALD2*, *aldA* and *aldB*.

Producing 3 - hydroxypropionic acid in a foreign host

In the work described below, the enzymes necessary for the production of 3 - hydroxypropionic acid from glycerol in *E. coli* were expressed under the regulation of the *trc* promoter, a non-native promoter inducible by the addition of IPTG. The glycerol dehydratase was encoded by the *dhaB* gene from *Klebsiella pneumoniae*, the aldehyde dehydrogenases used was any one of four different genes (*ALDH2* from *Homo sapiens*, *ALD4* from *S. cerevisiae*, *aldB* from *E. coli* or *aldA* from *E. coli*). Expression of these genes coding for glycerol dehydratase and any one of the genes encoding an aldehyde dehydrogenases was sufficient to enable the construct to produce 3-HP when the fermentation media was supplemented with glycerol. In all of these constructs, the *dhaB* gene was downstream from the gene encoding the aldehyde dehydrogenase used, and expression of both genes was regulated by the *trc* promoter. This order, however, is not required and the order of the gens on a construct and the use of multiple constructs is possible.

In a minimal genetic construct made based on the data presented here, the only genetic elements present that would be necessary are the structural genes *dhaB* and an aldehyde dehydrogenase gene encoding a protein that efficiently catalyzes the oxidation of 3-hydroxypropionaldehyde to 3-hydroxypropionic acid, and non-native promoter sequences specifically selected to give the type of inducible control most appropriate for the context of the process in which the construct is to be used. Extraneous pieces of DNA, whether retained in the construct or added from other DNA sequences, would not necessarily be detrimental to effective 3-HP synthesis by the host organism, but would not be needed. Each sequence to be translated would necessarily be preceded by a ribosome binding site, functional in the selected host so that the messenger RNA(s) coding for the proteins of interest could be translated by ribosomes. Terminator sequences immediately downstream of each translated unit would also be necessary in some organisms, particularly in eukaryotes. The construct could be part of an autonomously replicating sequence, such as a plasmid or phage vector, or could be

integrated into the genome of the host.

The structural genes and appropriate promoter(s) could be isolated by the use of restriction enzymes, by the polymerase chain reaction (PCR), by chemical synthesis of the appropriate oligonucleotides, or by other methods apparent to those skilled in the art or molecular biology. The promoter(s) would be derived from genomic DNA of other organism or from artificial genetic constructs containing promoters. Appropriate promoter fragments would be ligated into the construct upstream of the structural genes in any one of several possible arrangements.

The aldehyde dehydrogenase expressed would have: high specific activity towards 3-hydroxypropionaldehyde; be very stable in the host it is expressed in; be readily over expressed in the selected host; not be inhibited by either the substrates necessary for the reaction or the products formed by the reaction; be fully active under the fermentation conditions most favorable for the production of 3 - hydroxypropionic acid and be able to use either NAD⁺ or NADP⁺.

One possible arrangement is the true operon, where one promoter is used to direct transcription in one direction of all necessary Open Reading Frames (ORFs). The entire message is then contained in one messenger RNA. The advantages of the operon are that it is relatively easy to construct, since only one promoter is needed; that it is relatively simple to replace the promoter with another promoter if that would be desirable later; and that it assures that the two genes are under the same regulation. The main disadvantage of the operon scheme is that the levels of the expression of the two genes cannot be varied independently. If it is found that the genes, for optimal 3 - hydroxypropionic acid synthesis, should be expressed at different levels, the operon in most cases cannot be used to realize this.

Another possible arrangement is the multiple-promoter scheme. Two or more promoters, with the same or distinct regulatory behavior, could be used to direct transcription of the genes. For example, one promoter could be used to direct transcription of *dhaB* and one to direct transcription of the gene encoding the appropriate aldehyde dehydrogenases. Because the genes theoretically can be transcribed and translated separately, a great number of combinations of multiple promoters is possible. Additionally, it would be most desirable to prevent the promoters

from interfering with one another. This could be achieved either by placing two promoters into the construct such that they direct transcription in opposite directions, or by inserting transcriptional terminator sequences downstream of each separately transcribed unit. The main advantage of the multiple-promoter construct is that it permits independent regulation of as many distinct units as desired, which could be important. The disadvantages are that it would be more difficult to construct; more difficult to amend later; and more difficult to effectively regulate, since multiple changes in fermentation conditions would need to be introduced and might render the performance of the fermentation somewhat less predictable.

10 In any construct, the promoter sequence(s) used should be functional in the selected host organism and preferably provide sufficient transcription of the genes comprising the glycerol to 3 - hydroxypropionic acid pathway to enable the construct to be adequately active in that host. The promoter sequence(s) used would also effect regulation of transcription of the genes enabling the glycerol to 3-HP pathway to be
15 adequately active under the fermentation conditions employed for 3-HP production, and preferably they would be inducible, such that expression of the genes could be modulated by the inclusion in, or exclusion from, the fermentation of a certain agents or conditions.

A plausible example of the use of such a construct follows: one promoter, which
20 induced by the addition of an inexpensive chemical (the inducer) to the medium, could control transcription of both the *dhaB* gene and the gene encoding the appropriate aldehyde dehydrogenase. The cells would be permitted to grow in the absence of the inducer until they accumulated to a predetermined level. The inducer would then be added to the fermentation and nutritional changes commensurate with the altered
25 metabolism would be made to the medium as well. The cells would then be permitted to utilize the substrate(s) provided for 3-HP production (and additional biomass production if desired). After the cells could no longer use substrate to produce 3-HP, the fermentation would be stopped and the 3-HP recovered.

Genetic Sequences

30 To express glycerol dehydratase and a suitable aldehyde dehydrogenase, the two

enzymes necessary for the production of 3 - hydroxypropionic acid from glycerol, it is required that the DNA sequences containing the glycerol dehydratase and aldehyde dehydrogenase coding sequences be combined with at least a promoter sequence (preferably a non-native promoter although some native promoter activity may be present). An exemplary method of construction is described in the example below. To ensure that the present specification is enabling, the full sequences of the coding regions of genes for these enzymes is presented here.

Sequences 1, 3, 5 and 7 present different native genomic sequences for genes encoding aldehyde dehydrogenases.

10 SEQ ID NO:1 contains the full native DNA sequence encoding the *ALD4* enzyme from *Saccharomyces cerevisiae*. The amino acid sequence of the protein is presented as SEQ ID NO:2.

SEQ ID NO:3 includes the DNA sequence for the human *ALDH2* gene, again including the full protein coding region. The amino acid sequence for this human
15 alcohol dehydrogenase is presented in SEQ ID NO:4.

SEQ ID NO:5 and 7 respectively present the full coding sequences from the *E. coli* genes *aldA* and *aldB*, both of which encode alcohol dehydrogenases. The amino acid sequences for the proteins encoded by the genes are presented in SEQ ID NO: 6 and 8 respectively.

20 SEQ ID NO:9 contains the native genomic DNA sequence for the *dhaB* gene from the *dha* regulon of *Klebsiella pneumoniae*. The coding sequences for this complex regulon produces five polypeptides, which are presented as SEQ ID NOS:10 through 13, which together provide the activity of the glycerol dehydratase enzyme.

Each of these coding sequences can be used to make genetic constructs for the
25 expression of the appropriate enzymes in a heterologous hosts. In making genetic constructs for expression of the genes in such hosts, it is contemplated that heterologous promoters will be joined to the coding sequences for the enzymes, but all that it required is that the promoters be effective for the hosts in which the genes are to be expressed. It is also contemplated and envisioned that significant variations in DNA sequence are
30 possible from the native DNA coding sequences presented here. As is well known in the art, due to the degeneracy of the genetic code, many different DNA sequences can

encode the expression of the same protein. So, when this document uses language specifying a DNA sequence encoding a protein, it is intended to encompass any DNA sequence which can be used to express that protein even if different from the genomic sequences presented here. It is also contemplated that conservative changes in the amino acid sequences of the proteins specified here can be made without departing from the present invention. In particular, deletions, additions and substitutions of one or more amino acids in a protein sequence can almost always be made without changing protein functionality. When the name of a protein is used here, it is intended to be equally applicable to both such minor changes in amino acid sequence and to allelic variations in native protein sequence as occurs within the species named as well as other closely related species.

It is possible that many of the above DNA sequences could be truncated and still express a protein that has the same enzymatic properties. One skilled in the art of molecular biology would appreciate that minor deletions, additions and mutations may not change the attributes of the designated base pair sequences; many of the nucleotide of the designated base pair sequences are probably not essential for their unique function. To determine whether or not an altered sequence or sequences has sufficient homology with the designated base pairs to function identically, one would simply create the candidate mutation, deletion or alteration and create a gene construct including the altered sequence together with promoter and termination sequences. This gene construct could be tested as, described below, for the production of 3-HP from glycerol.

Certain DNA primers were used to isolate or clone the genomic DNA sequences used in the experiments described below. While the sequence information presented here is sufficient to enable the construction of expression plasmids incorporating the genes identified here, in order to redundantly enable the use of these genes, primers which may be used to isolate the genes from their native hosts are described below.

The primers aldA_L (SEQ ID NO:14), and aldA_R (SEQ ID NO:15), were used to amplify the 1513 bp *aldA* fragment from genomic *E. coli* DNA (strain MG1655, a gift from the Genetic Stock Center, New Haven, CT). The gel purified PCR fragment containing a DNA sequence coding for the expression of aldehyde dehydrogenase was

inserted into *NcoI-XhoI* site of pSE380 (Invitrogen, San Diego, CA) to give pPFS3. The resulting plasmid contained *aldA* under the control of the *trc* promoter. This construct allowed for high-level expression of the *aldA* gene from *E. coli* under regulation of the *trc* promoter. Unless indicated otherwise all molecular biology and plasmid

5 constructions were done in *E. coli* AG1 (Stratagene, La Jolla, CA).

The primers aldB_L (SEQ ID NO:20) and aldB_R (SEQ ID NO:21), were used to amplify the 1574 bp *aldB* fragment from genomic *E. coli* DNA (strain MG1655). The resulting PCR converted the TGA stop codon into a TAA stop codon. The gel-purified PCR fragment containing the DNA sequence sufficiently coding for the
10 expression of aldehyde dehydrogenase was inserted into the *KpnI-SacI* site of pSE380 to give pPFS12.

The primers ALD4_L (SEQ ID NO : 16), and ALD4_R (SEQ ID NO : 17), were used to amplify the 1595 bp ALD4 fragment from *S. cerevisiae* DNA (strain YPH500). The gel-purified fragment containing a DNA sequence coding for the expression of
15 aldehyde dehydrogenase was inserted into the *KpnI-SacI* site of pPFS3 to give pPFS8. The resulting plasmid contained mature *ALD4* under control of the *trc* promoter.

The primers ALDH2_L (SEQ ID NO:18), and ALDH2_R (SEQ ID NO:19), were used to amplify the 1541 bp ALDH2 fragment from pT7-7::ALDH2, a gift from H. Weiner (Purdue University, West Lafayette, IN). The gel purified PCR fragment
20 containing a DNA sequence sufficiently homologous to base pairs 22 to 1524, inclusive of SEQ ID NO : 3 so as to code for the expression of aldehyde dehydrogenase was inserted in to the *KpnI-SacI* site of pSE380 to give pPFS7. This sequence was moved from pPFS7 into the *KpnI-SacI* site of pPFS3 to give pPFS9. The resulting plasmid contained mature ALDH2 under the control of the *trc* promoter.

25 The primers pTRC_L (SEQ ID NO:22), and pTRC_R (SEQ ID NO:23), were used to amplify the 540 bp fragment from pSE380. The gel purified PCR fragment was inserted into the *HpaI-KpnI* site of pPFS3 to give pPFS13. The resulting plasmid deleted the "native" ribosome binding site of pSE380 and a *NcoI* site (which contained an extraneous ATG start codon upstream of the cloned genes). The *KpnI-SacI*
30 fragments of pPFS8, pPFS9, and pPFS12 were inserted into the *KpnI-SacI* site of pPFS13 to give pPFS14, pPFS15, and pPFS16, respectively.

Assay for production of 3-HP

The efficacy of changes made as contemplated herein can be checked by the following tests. To test for the production of 3-HP, fermentation products can be quantified with a Waters Alliance Integrity HPLC system (Milford, MA) equipped with a refractive index detector, a photodiode array detector, and an Aminex HPX-87H (Bio-Rad, Hercules, CA) organic acids column. The mobile phase should be 0.01 N sulfuric acid solution (pH 2.0) at a flow rate of 0.5 mL/min. The column temperature should be set to 40°C. Compounds can be identified by determining if they co-elute with authentic standards. Prior to analysis, all samples should be filtered through 0.45 µM pore size membrane. (Gelman Sciences, Ann Arbor, MI). The fractions of the fermentation products collected using HPLC should be analyzed on a Varian Star 3400 CX, gas - chromatograph coupled to a Varian Saturn 3 mass spectrometer (GC-MS) (Walnut Creek, CA).

Assay for enzyme activity.

Aldehyde dehydrogenase activity can be determined by measuring the reduction of β -NAD⁺ at 25°C with 3 - hydroxypropionaldehyde as a substrate. All buffers should contain 1 mM ethylenediaminetetraacetic acid (EDTA), 0.1 mM Pefabloc SC (Boehringer Mannheim, Indianapolis, IN) and 1 mM Tris (carboxyethyl) phosphine hydrochloride (TCEP-HCL). For ALD4, the solution should contain 100 mM Tris HCL Buffer (pH 8.0), 100 mM KCl. For ALDH2 the solution should contained 100 mM sodium pyrophosphate (pH 9.0). For AldA and AldB, the solution should contain 20 mM sodium glycine (pH 9.5). A total of 3.0 mL of buffer should be added to quartz cuvettes and allowed to equilibrate to assay temperature. From 5 to 20 µL of cell extract should be added and background activity recorded after the addition of β -NAD⁺ to a final concentration of 0.67 mM. The reaction should be started by the addition of substrate (either acetaldehyde, propionaldehyde, or 3 - hydroxypropionaldehyde) to a final concentration of 2 mM. Assay mixtures should be stirred with micro-stirrers during the assays.

For aldehyde dehydrogenase activity assays, one unit is defined as the reduction

of 1.0 μ M of β -AND⁺ per minute at 25° C. These reactions can be monitored by following the change in absorbance at 340 nm (A_{340}) at 25°C on a Varain Carry-1 Bio spectrophotometer (Sugar Land, TX). Total protein concentrations in the cell extracts can be determined using the Bradford assay method (Bio-Rad, Hercules, CA) with
5 bovine serum albumin as the standard.

EXAMPLES

Plasmid constructions.

Klebsiella pneumoniae expresses glycerol dehydratase, an enzyme that catalyzes the conversion of glycerol to 3 - hydroxypropionaldehyde, (*dhaB*) and 1,3 -
10 propanediol oxidoreductase an enzyme that catalyzes the conversion of 3 - hydroxypropionaldehyde to 1,3 - propanediol respectively (the gene product from *dhaT*). A plasmid encoding these two genes was created and expressed in *E. coli* (plasmid pTC53). The *dhaT* gene was deleted from pTC53 to create pMH34. The resulting plasmid still contained the DNA sequence complementary to base pairs 330 to
15 2153 inclusion of SEQ ID NO : 9, the complement of base pairs 2166 to 2591, inclusive, of SEQ ID NO : 9, and the complement of base pairs 3191 to 4858, inclusive, of SEQ ID NO : 9, so as to code for the expression of glycerol dehydratase. The fragment of DNA encoding these sequences was excised from pMH34 by cutting it with *Sall-XbaI*, and the resulting fragment was gel purified (the purified fragment was gift
20 from M. Hoffman of the University of Wisconsin - Madison). This DNA fragment was inserted into the *Sall-XbaI* site of pPFS13 to give pPFS17.

The resulting plasmid contained both the *aldA* and *dhaB* genes under the control of the *trc* promoter. Similarly, the gel-purified *Sall-XbaI* fragment from pMH34 was inserted into the *Sall-XbaI* sites of pPFS14, pPFS15, and pPFS16 to give pPFS18,
25 pPFS19, and pPFS20, respectively. These plasmids contained *ALD4*, *ALDH2*, and *aldB*, respectively, as well as *dhaB* under the control of the *trc* promoter; in all of the constructs the *dhaB* gene were downstream of the gene encoding the aldehyde dehydrogenase.

Expression in *E. coli*.

The efficacy of *E. coli* as a platform for the production of 3-HP from growth on glucose has been examined using a mathematical model developed for this purpose. The model was executed in two different ways assuming the conversion of one mole of
5 glucose under either anaerobic or aerobic conditions either directly to 3-HP or to the production of 3-HP and ATP. The optimum yield under anaerobic conditions is 1 mole of 3-HP and 1 mole of lactate. The more realistic yield under anaerobic conditions is 0.5 moles of 3-HP, 1.5 moles of lactate and 1 mole of ATP. The optimum yield under aerobic conditions is 1.9 moles of 3-HP and 0.3 moles of CO₂. The realistic yield under
10 aerobic conditions is 1.85 moles of 3-HP, 0.35 moles of CO₂ and 1 mole of ATP.

The effect of 3-HP concentration on *E. coli* strain MG1655 growth was measured. Cells were grown on standard media with and without the addition of up to 80g/L of 3-HP. The best fit of these data demonstrated that 3-HP was only 1.4 times as inhibitory as lactic acid on the growth of *E. coli*. It is possible to economically produce
15 lactic acid using *E. coli*, since 3-HP is only 1.4 times more inhibitory than lactic acid, it should be possible to use *E. coli* as a host for the commercial production of 3-HP.

Media and growth conditions

The standard media contained the following per liter: 6 g Na₂HPO₄, 3 g KH₂PO₄, 1 g NH₄Cl, 0.5 g NaCl, 3 mg CaCl₂, 5 g yeast extract (Difco Laboratories, Detroit, MI)
20 and 2 mM MgSO₄. When necessary to retain plasmids ampicillin (100 mg/mL) was added to the media. Isopropyl-β-thiogalactopyranoside (IPTG) was added in varying amounts to induce gene expression. All fermentations were carried out in an incubator-shaker at 37 C and 200 rpm. Anaerobic fermentations were carried out in 500-mL anaerobic flasks with 300 mL of working volume. Inocula for fermentations were
25 grown overnight in Luria-Bertani medium supplemented with ampicillin is necessary. The 300-mL fermentations were inoculated with 1.5 mL of the overnight culture. For enzyme assays, fermentations were incubated for 24 hours.

Over expression of aldehyde dehydrogenase in *E. coli*.

Cells were harvested by centrifugation at 3000 x g for 10 minutes at 4°C with a

Beckman (Fullerton, CA) model J2-21 centrifuge. Cell pellets were washed twice in 100 mM potassium phosphate buffer at pH 7.2 and re-suspended in appropriate assay resuspension buffer equal to 5 x of the volume of the wet cell mass. The cells were homogenized using a French pressure cell. The homogenate was centrifuged at 40000 x g for 30 minutes. The supernatant was dialyzed against the appropriate resuspension buffer using 10000 molecular weight cut-off pleated dialysis tubing (Pierce, Rockford, IL) at 4°C. Dialysis buffer was changed after 2 hours, and 4 hours, and dialysis was stopped after being allowed to proceed overnight.

E. coli AG1 cells transfected with the plasmids constructed to express the *aldA*, *ALD4*, *ALDH2*, or *aldB* genes were grown in 500-mL anaerobic flasks. Twelve hours after the fermentations were inoculated IPTG was added to induce enzyme expression. The cells were allowed to grow for an additional 12 hours then harvested and lysed as discussed above. The soluble fraction of the lysate was assayed for aldehyde dehydrogenase activity using the substrate 3-hydroxypropionaldehyde in the buffer appropriate for the particular enzyme expressed. The plasmid, aldehyde dehydrogenase expressed and specific activity measured (U/mg of protein) were as follows: pPFS13, *aldA*, 0.2; pPFS14, *ALD4*, 0.5, pPFS15, *ALDH2*, 0.3; and pPFS16, *aldB*, 0.1. The control, *E. coli* strain AG1 harboring plasmid pSE380, encoded no exogenous aldehyde dehydrogenase activity and it had no detectable activity with 3-HP as substrate. It is clear from the activity assays that all four aldehyde dehydrogenases were expressed in *E. coli*. The aldehyde dehydrogenase cloned from *Saccharomyces cerevisiae* (*ADH4*) had the highest activity when 3-hydroxypropionaldehyde was used as the substrate (0.5 units/mg of protein).

E. coli cells transformed with plasmids expressing: aldehyde dehydrogenase; both aldehyde dehydrogenase and glycerol dehydratase, or neither gene; were grown and assayed for their ability to produce 3-HP from glycerol. The cells were grown on standard media supplemented with 6 µM of Coenzyme B₁₂, under anaerobic conditions in the absence of light (to protect the integrity of the Coenzyme B₁₂ necessary for DhaB activity). After 12 hours, IPTG was added to induce expression of the genes under the *trc* promoter at the same time 5g/L of glycerol was added. After 12 more hours of anaerobic fermentation the fermentation broth was assayed for 3 - HP by HPLC and GC,

the plasmid, aldehyde dehydrogenase gene expressed and g/L of 3- HP measured were as follows: pSF17, *aldA*, 0.031; pPSF18 *ALD4*, 0.173; and pPSF19, *ALDH2*, 0.061.

Cells expressing *dhaB* but no exogenous aldehyde dehydrogenase genes (plasmid pMH34) produced 0.015 g/L of 3 - HP. Cells expressing *aldA*, *ALD4*, *ALDH2* or *aldB*
5 but not *dhaB* (plasmids pPFS13, pPFS14, pPFS15, pPFS16, respectively) all produced less then 0.005 g/L of 3-HP when the media the cells were growing in was supplemented with 2.5g/L of 3-hydroxypropionaldehyde.

Other Hosts and Promoters

Applications of the 3 - hydroxypropionic acid pathway such as the genetic
10 constructs of the present invention can easily be expressed in other organisms. The required genes would need to be placed under control of an appropriate promoter or promoters. Some organism such as yeasts may require transcription terminators to be placed after each transcribed unit. The knowledge of the present intention makes such amendments possible. Such a genetic construct would need to be part of a vector that
15 could either replicate in the new host or integrate into the chromosome of the new host. Many such vectors are commercially available for expression in gram-negative and gram-positive bacteria, yeast, mammalian cells, insect cell, plant, etc. For example, to express the 3-hydroxypropionic acid pathway in *Rhodobacter capsulatus*, one could obtain vector pNH2 from the American Type Culture Collection (ATTC). This is a
20 shuttle vector for use in *R. capsulatus* and *E. coli*. Organisms such as *Saccharomyces cerevisiae* which can convert glucose to glycerol could be used as a host, such a construct would enable the production of 3 - HP directly from glucose. Additionally, other substrates such as xylan could also be used given the selection of an appropriate host.

25 Stochiometric analysis shows that best stochiometric yield of 3-HP production in *E. coli* calculated on the basis of glucose consumed is obtained under aerobic conditions. Under aerobic condition CO₂ is the only carbon-containing co-product, in particular the generation of lactic acid which occurs under anaerobic conditions is avoided. Production of 3-HP under these conditions could result in a more economical
30 recovery of 3-HP from the fermentation broth.

Alternatively, the *dhaB* gene and a gene encoding the appropriate aldehyde dehydrogenase could be cloned into the multiple cloning site of this vector in *E. coli* to facilitate construction, and then transformed into *R. capsulatus*. The *R. capsulatus* *nifH* promoter, provided on the plasmid, could be used to direct the transcription in *R.*

- 5 *capsulatus* of the genes placed into pNF2 in series with one promoter, or with two copies of the *nifH* promoter. Expression of the genes in other organisms would require a procedure analogous to that presented here.

Alternative Aldehyde Dehydrogenases and Glycerol Dehydratases

- Applications of the pathway for the production of 3-hydroxypropionic acid from
10 glycerol can be made using other suitable aldehyde dehydrogenases. To be functional in this pathway an aldehyde dehydrogenase needs to be stable, readily expressed in the host of choice and have high enough activity towards 3-hydroxypropionaldehyde to enable it to make 3-HP. The knowledge of the present invention makes such amendments possible. A program of directed evolution could be undertaken to select
15 for suitable aldehyde dehydrogenases or they could be recovered from native sources, the genes encoding these enzymes in conjunction with a gene encoding an appropriate glycerol dehydratase activity, would then be made part of any of the constructs envisioned here to produce 3 - hydroxypropionic acid from glycerol.

- A similar program of enzyme improvement including for example directed
20 evolution could be carried out using the *dhaB* gene from *Klebsiella pneumoniae* as a starting point to obtain other variants of glycerol dehydratase that are superior in efficiency and stability to the form used in this invention. Alternatively, enzymes which catalyzes the same reaction may be isolated from others organisms and used in place of the *Klebsiella pneumoniae* glycerol dehydratase. Such enzymes may be especially
25 useful in alternative hosts wherein they may be more readily expressed, be more stable and more efficient under the fermentation conditions best suited to the growth of the construct and the production and recovery of 3-HP.

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Gln Glu Gly Ala Lys Leu Leu Cys Gly Gly Gly Ile Ala Ala Asp Arg	
365 370 375	
ggt tac ttc atc cag ccc act gtg ttt gga gat gtg cag gat ggc atg	1203
Gly Tyr Phe Ile Gln Pro Thr Val Phe Gly Asp Val Gln Asp Gly Met	
380 385 390	
acc atc gcc aag gag gag atc ttc ggg cca gtg atg cag atc ctg aag	1251
Thr Ile Ala Lys Glu Glu Ile Phe Gly Pro Val Met Gln Ile Leu Lys	
395 400 405 410	
ttc aag acc ata gag gag gtt gtt ggg aga gcc aac aat tcc acg tac	1299
Phe Lys Thr Ile Glu Glu Val Val Gly Arg Ala Asn Asn Ser Thr Tyr	
415 420 425	
ggg ctg gcc gca gct gtc ttc aca aag gat ttg gac aag gcc aat tac	1347
Gly Leu Ala Ala Ala Val Phe Thr Lys Asp Leu Asp Lys Ala Asn Tyr	
430 435 440	

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ctg tcc cag gcc ctc cag gcg ggc act gtg tgg gtc aac tgc tat gat 1395
Leu Ser Gln Ala Leu Gln Ala Gly Thr Val Trp Val Asn Cys Tyr Asp
445 450 455

gtg ttt gga gcc cag tca ccc ttt ggt ggc tac aag atg tcg ggg agt 1443
Val Phe Gly Ala Gln Ser Pro Phe Gly Gly Tyr Lys Met Ser Gly Ser
460 465 470

ggc cgg gag ttg ggc gag tac ggg ctg cag gca tac act gaa gtg aaa 1491
Gly Arg Glu Leu Gly Glu Tyr Gly Leu Gln Ala Tyr Thr Glu Val Lys
475 480 485 490

act gtc aca gtc aaa gtg cct cag aag aac tcataagagc tcgaattcgc 1541
Thr Val Thr Val Lys Val Pro Gln Lys Asn
495 500

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<211> 500

<212> PRT

<213> Homo sapiens

<400> 4

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20 25 30

Val Ser Arg Lys Thr Phe Pro Thr Val Asn Pro Ser Thr Gly Glu Val
35 40 45

Ile Cys Gln Val Ala Glu Gly Asp Lys Glu Asp Val Asp Lys Ala Arg
50 55 60

Glu Gly Arg Pro Gly Ala Phe Gln Leu Gly Ser Pro Trp Arg Arg Met
65 70 75 80

Asp Ala Ser His Ser Gly Arg Leu Leu Asn Arg Leu Ala Asp Leu Ile
85 90 95

Glu Arg Asp Arg Thr Tyr Leu Ala Ala Leu Glu Thr Leu Asp Asn Gly
100 105 110

Lys Pro Tyr Val Ile Ser Tyr Leu Val Asp Leu Asp Met Val Leu Lys
115 120 125

Cys	Leu	Arg	Tyr	Tyr	Ala	Gly	Trp	Ala	Asp	Lys	Tyr	His	Gly	Lys	Thr
130						135						140			
Ile	Pro	Ile	Asp	Gly	Asp	Phe	Phe	Ser	Tyr	Thr	Arg	His	Glu	Pro	Val
145					150					155					160
Gly	Val	Cys	Gly	Gln	Ile	Ile	Pro	Trp	Asn	Phe	Pro	Leu	Leu	Met	Gln
				165					170					175	
Ala	Trp	Lys	Leu	Gly	Pro	Ala	Leu	Ala	Thr	Gly	Asn	Val	Val	Val	Met
			180					185					190		
Lys	Val	Ala	Glu	Gln	Thr	Pro	Leu	Thr	Ala	Leu	Tyr	Val	Ala	Asn	Leu
		195					200					205			
Ile	Lys	Glu	Ala	Gly	Phe	Pro	Pro	Gly	Val	Val	Asn	Ile	Val	Pro	Gly
	210					215					220				
Phe	Gly	Pro	Thr	Ala	Gly	Ala	Ala	Ile	Ala	Ser	His	Glu	Asp	Val	Asp
225					230					235					240
Lys	Val	Ala	Phe	Thr	Gly	Ser	Thr	Glu	Ile	Gly	Arg	Val	Ile	Gln	Val
				245					250					255	
Ala	Ala	Gly	Ser	Ser	Asn	Leu	Lys	Arg	Val	Thr	Leu	Glu	Leu	Gly	Gly
			260					265					270		
Lys	Ser	Pro	Asn	Ile	Ile	Met	Ser	Asp	Ala	Asp	Met	Asp	Trp	Ala	Val
		275					280					285			
Glu	Gln	Ala	His	Phe	Ala	Leu	Phe	Phe	Asn	Gln	Gly	Gln	Cys	Cys	Cys
	290					295					300				
Ala	Gly	Ser	Arg	Thr	Phe	Val	Gln	Glu	Asp	Ile	Tyr	Asp	Glu	Phe	Val
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Val	Arg	Ser	Val	Ala	Arg	Ala	Lys	Ser	Arg	Val	Val	Gly	Asn	Pro	Phe
				325					330					335	
Asp	Ser	Lys	Thr	Glu	Gln	Gly	Pro	Gln	Val	Asp	Glu	Thr	Gln	Phe	Lys
			340					345					350		
Lys	Ile	Leu	Gly	Tyr	Ile	Asn	Thr	Gly	Lys	Gln	Glu	Gly	Ala	Lys	Leu
		355					360					365			
Leu	Cys	Gly	Gly	Gly	Ile	Ala	Ala	Asp	Arg	Gly	Tyr	Phe	Ile	Gln	Pro
	370					375					380				

Thr Val Phe Gly Asp Val Gln Asp Gly Met Thr Ile Ala Lys Glu Glu
385 390 395 400

Ile Phe Gly Pro Val Met Gln Ile Leu Lys Phe Lys Thr Ile Glu Glu
405 410 415

Val Val Gly Arg Ala Asn Asn Ser Thr Tyr Gly Leu Ala Ala Val
420 425 430

Phe Thr Lys Asp Leu Asp Lys Ala Asn Tyr Leu Ser Gln Ala Leu Gln
435 440 445

Ala Gly Thr Val Trp Val Asn Cys Tyr Asp Val Phe Gly Ala Gln Ser
450 455 460

Pro Phe Gly Gly Tyr Lys Met Ser Gly Ser Gly Arg Glu Leu Gly Glu
465 470 475 480

Tyr Gly Leu Gln Ala Tyr Thr Glu Val Lys Thr Val Thr Val Lys Val
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Pro Gln Lys Asn
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<213> Escherichia coli

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His Pro Met Tyr Ile Asp Gly Gln Phe Val Thr Trp Arg Gly Asp Ala
10 15 20

tgg att gat gtg gta aac cct gct aca gag gct gtc att tcc cgc ata 150
Trp Ile Asp Val Val Asn Pro Ala Thr Glu Ala Val Ile Ser Arg Ile
25 30 35

ccc gat ggt cag gcc gag gat gcc cgt aag gca atc gat gca gca gaa 198
 Pro Asp Gly Gln Ala Glu Asp Ala Arg Lys Ala Ile Asp Ala Ala Glu
 40 45 50

cgt gca caa cca gaa tgg gaa gcg ttg cct gct att gaa cgc gcc agt 246
 Arg Ala Gln Pro Glu Trp Glu Ala Leu Pro Ala Ile Glu Arg Ala Ser
 55 60 65 70

tgg ttg cgc aaa atc tcc gcc ggg atc cgc gaa cgc gcc agt gaa atc 294
 Trp Leu Arg Lys Ile Ser Ala Gly Ile Arg Glu Arg Ala Ser Glu Ile
 75 80 85

agt gcg ctg att gtt gaa gaa ggg ggc aag atc cag cag ctg gct gaa 342
 Ser Ala Leu Ile Val Glu Glu Gly Gly Lys Ile Gln Gln Leu Ala Glu
 90 95 100

gtc gaa gtg gct ttt act gcc gac tat atc gat tac atg gcg gag tgg 390
 Val Glu Val Ala Phe Thr Ala Asp Tyr Ile Asp Tyr Met Ala Glu Trp
 105 110 115

gca cgg cgt tac gag ggc gag att att caa agc gat cgt cca gga gaa 438
 Ala Arg Arg Tyr Glu Gly Glu Ile Ile Gln Ser Asp Arg Pro Gly Glu
 120 125 130

aat att ctt ttg ttt aaa cgt gcg ctt ggt gtg act acc ggc att ctg 486
 Asn Ile Leu Leu Phe Lys Arg Ala Leu Gly Val Thr Thr Gly Ile Leu
 135 140 145 150

ccg tgg aac ttc ccg ttc ttc ctc att gcc cgc aaa atg gct ccc gct 534
 Pro Trp Asn Phe Pro Phe Phe Leu Ile Ala Arg Lys Met Ala Pro Ala
 155 160 165

ctt ttg acc ggt aat acc atc gtc att aaa cct agt gaa ttt acg aca 582
 Leu Leu Thr Gly Asn Thr Ile Val Ile Lys Pro Ser Glu Phe Thr Thr
 170 175 180

aac aat gcg att gca ttc gcc aaa atc gtc gat gaa ata ggc ctt ccg 630
 Asn Asn Ala Ile Ala Phe Ala Lys Ile Val Asp Glu Ile Gly Leu Pro
 185 190 195

cgc ggc gtg ttt aac ctt gta ctg ggg cgt ggt gaa acc gtt ggg caa 678
 Arg Gly Val Phe Asn Leu Val Leu Gly Arg Gly Glu Thr Val Gly Gln
 200 205 210

gaa ctg gcg ggt aac cca aag gtc gca atg gtc agt atg aca ggc agc 726
 Glu Leu Ala Gly Asn Pro Lys Val Ala Met Val Ser Met Thr Gly Ser
 215 220 225 230

gtc tct gca ggt gag aag atc atg gcg act gcg gcg aaa aac atc acc	774
Val Ser Ala Gly Glu Lys Ile Met Ala Thr Ala Ala Lys Asn Ile Thr	
235 240 245	
aaa gtg tgt ctg gaa ttg ggg ggt aaa gca cca gct atc gta atg gac	822
Lys Val Cys Leu Glu Leu Gly Gly Lys Ala Pro Ala Ile Val Met Asp	
250 255 260	
gat gcc gat ctt gaa ctg gca gtc aaa gcc atc gtt gat tca cgc gtc	870
Asp Ala Asp Leu Glu Leu Ala Val Lys Ala Ile Val Asp Ser Arg Val	
265 270 275	
att aat agt ggg caa gtg tgt aac tgt gca gaa cgt gtt tat gta cag	918
Ile Asn Ser Gly Gln Val Cys Asn Cys Ala Glu Arg Val Tyr Val Gln	
280 285 290	
aaa ggc att tat gat cag ttc gtc aat cgg ctg ggt gaa gcg atg cag	966
Lys Gly Ile Tyr Asp Gln Phe Val Asn Arg Leu Gly Glu Ala Met Gln	
295 300 305 310	
gcg gtt caa ttt ggt aac ccc gct gaa cgc aac gac att gcg atg ggg	1014
Ala Val Gln Phe Gly Asn Pro Ala Glu Arg Asn Asp Ile Ala Met Gly	
315 320 325	
ccg ttg att aac gcc gcg gcg ctg gaa agg gtc gag caa aaa gtg gcg	1062
Pro Leu Ile Asn Ala Ala Ala Leu Glu Arg Val Glu Gln Lys Val Ala	
330 335 340	
cgc gca gta gaa gaa ggg gcg aga gtg gcg ttc ggt ggc aaa gcg gta	1110
Arg Ala Val Glu Glu Gly Ala Arg Val Ala Phe Gly Gly Lys Ala Val	
345 350 355	
gag ggg aaa gga tat tat tat ccg ccg aca ttg ctg ctg gat gtt cgc	1158
Glu Gly Lys Gly Tyr Tyr Tyr Pro Pro Thr Leu Leu Leu Asp Val Arg	
360 365 370	
cag gaa atg tcg att atg cat gag gaa acc ttt ggc ccg gtg ctg cca	1206
Gln Glu Met Ser Ile Met His Glu Glu Thr Phe Gly Pro Val Leu Pro	
375 380 385 390	
gtt gtc gca ttt gac acg ctg gaa gat gct atc tca atg gct aat gac	1254
Val Val Ala Phe Asp Thr Leu Glu Asp Ala Ile Ser Met Ala Asn Asp	
395 400 405	
agt gat tac ggc ctg acc tca tca atc tat acc caa aat ctg aac gtc	1302
Ser Asp Tyr Gly Leu Thr Ser Ser Ile Tyr Thr Gln Asn Leu Asn Val	
410 415 420	

11 01 03 30 25 11 01 03 00 00

gcg atg aaa gcc att aaa ggg ctg aag ttt ggt gaa act tac atc aac 1350
Ala Met Lys Ala Ile Lys Gly Leu Lys Phe Gly Glu Thr Tyr Ile Asn
425 430 435

cgt gaa aac ttc gaa gct atg caa ggc ttc cac gcc gga tgg cgt aaa 1398
Arg Glu Asn Phe Glu Ala Met Gln Gly Phe His Ala Gly Trp Arg Lys
440 445 450

tcc ggt att ggc ggc gca gat ggt aaa cat ggc ttg cat gga tat ctg 1446
Ser Gly Ile Gly Gly Ala Asp Gly Lys His Gly Leu His Gly Tyr Leu
455 460 465 470

cag acc cag gtg gtt tat tta cag tct taagagctcg aattcccgtc 1493
Gln Thr Gln Val Val Tyr Leu Gln Ser
475

gacggctcta gactcgagcg 1513

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<212> PRT
<213> Escherichia coli
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20 25 30

Ala Val Ile Ser Arg Ile Pro Asp Gly Gln Ala Glu Asp Ala Arg Lys
35 40 45

Ala Ile Asp Ala Ala Glu Arg Ala Gln Pro Glu Trp Glu Ala Leu Pro
50 55 60

Ala Ile Glu Arg Ala Ser Trp Leu Arg Lys Ile Ser Ala Gly Ile Arg
65 70 75 80

Glu Arg Ala Ser Glu Ile Ser Ala Leu Ile Val Glu Glu Gly Gly Lys
85 90 95

Ile Gln Gln Leu Ala Glu Val Glu Val Ala Phe Thr Ala Asp Tyr Ile
100 105 110

Asp Tyr Met Ala Glu Trp Ala Arg Arg Tyr Glu Gly Glu Ile Ile Gln
115 120 125

Ser Asp Arg Pro Gly Glu Asn Ile Leu Leu Phe Lys Arg Ala Leu Gly
 130 135 140

Val Thr Thr Gly Ile Leu Pro Trp Asn Phe Pro Phe Phe Leu Ile Ala
 145 150 155 160

Arg Lys Met Ala Pro Ala Leu Leu Thr Gly Asn Thr Ile Val Ile Lys
 165 170 175

Pro Ser Glu Phe Thr Thr Asn Asn Ala Ile Ala Phe Ala Lys Ile Val
 180 185 190

Asp Glu Ile Gly Leu Pro Arg Gly Val Phe Asn Leu Val Leu Gly Arg
 195 200 205

Gly Glu Thr Val Gly Gln Glu Leu Ala Gly Asn Pro Lys Val Ala Met
 210 215 220

Val Ser Met Thr Gly Ser Val Ser Ala Gly Glu Lys Ile Met Ala Thr
 225 230 235 240

Ala Ala Lys Asn Ile Thr Lys Val Cys Leu Glu Leu Gly Gly Lys Ala
 245 250 255

Pro Ala Ile Val Met Asp Asp Ala Asp Leu Glu Leu Ala Val Lys Ala
 260 265 270

Ile Val Asp Ser Arg Val Ile Asn Ser Gly Gln Val Cys Asn Cys Ala
 275 280 285

Glu Arg Val Tyr Val Gln Lys Gly Ile Tyr Asp Gln Phe Val Asn Arg
 290 295 300

Leu Gly Glu Ala Met Gln Ala Val Gln Phe Gly Asn Pro Ala Glu Arg
 305 310 315 320

Asn Asp Ile Ala Met Gly Pro Leu Ile Asn Ala Ala Ala Leu Glu Arg
 325 330 335

Val Glu Gln Lys Val Ala Arg Ala Val Glu Glu Gly Ala Arg Val Ala
 340 345 350

Phe Gly Gly Lys Ala Val Glu Gly Lys Gly Tyr Tyr Tyr Pro Pro Thr
 355 360 365

Leu Leu Leu Asp Val Arg Gln Glu Met Ser Ile Met His Glu Glu Thr
 370 375 380

Phe Gly Pro Val Leu Pro Val Val Ala Phe Asp Thr Leu Glu Asp Ala
385 390 395 400

Ile Ser Met Ala Asn Asp Ser Asp Tyr Gly Leu Thr Ser Ser Ile Tyr
405 410 415

Thr Gln Asn Leu Asn Val Ala Met Lys Ala Ile Lys Gly Leu Lys Phe
420 425 430

Gly Glu Thr Tyr Ile Asn Arg Glu Asn Phe Glu Ala Met Gln Gly Phe
435 440 445

His Ala Gly Trp Arg Lys Ser Gly Ile Gly Gly Ala Asp Gly Lys His
450 455 460

Gly Leu His Gly Tyr Leu Gln Thr Gln Val Val Tyr Leu Gln Ser
465 470 475

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<211> 1574
<212> DNA
<213> Escherichia coli

<220>
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<222> (22)..(1557)

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1 5 10
aag ccc ggc gag tat ggt ttc ccc ctc aag tta aaa gcc cgc tat gac 99
Lys Pro Gly Glu Tyr Gly Phe Pro Leu Lys Leu Lys Ala Arg Tyr Asp
15 20 25
aac ttt att ggc ggc gaa tgg gta gcc cct gcc gac ggc gag tat tac 147
Asn Phe Ile Gly Gly Glu Trp Val Ala Pro Ala Asp Gly Glu Tyr Tyr
30 35 40
cag aat ctg acg ccg gtg acc ggg cag ctg ctg tgc gaa gtg gcg tct 195
Gln Asn Leu Thr Pro Val Thr Gly Gln Leu Leu Cys Glu Val Ala Ser
45 50 55
tcg ggc aaa cga gac atc gat ctg gcg ctg gat gct gcg cac aaa gtg 243

Ser	Gly	Lys	Arg	Asp	Ile	Asp	Leu	Ala	Leu	Asp	Ala	Ala	His	Lys	Val	
60						65					70					
aaa	gat	aaa	tgg	gcg	cac	acc	tcg	gtg	cag	gat	cgt	gcg	gcg	att	ctg	291
Lys	Asp	Lys	Trp	Ala	His	Thr	Ser	Val	Gln	Asp	Arg	Ala	Ala	Ile	Leu	
75					80					85					90	
ttt	aag	att	gcc	gat	cga	atg	gaa	caa	aac	ctc	gag	ctg	tta	gcg	aca	339
Phe	Lys	Ile	Ala	Asp	Arg	Met	Glu	Gln	Asn	Leu	Glu	Leu	Leu	Ala	Thr	
					95					100					105	
gct	gaa	acc	tgg	gat	aac	ggc	aaa	ccc	att	cgc	gaa	acc	agt	gct	gcg	387
Ala	Glu	Thr	Trp	Asp	Asn	Gly	Lys	Pro	Ile	Arg	Glu	Thr	Ser	Ala	Ala	
			110					115						120		
gat	gta	ccg	ctg	gcg	att	gac	cat	ttc	cgc	tat	ttc	gcc	tcg	tgt	att	435
Asp	Val	Pro	Leu	Ala	Ile	Asp	His	Phe	Arg	Tyr	Phe	Ala	Ser	Cys	Ile	
			125					130						135		
cgg	gcg	cag	gaa	ggc	ggg	atc	agt	gaa	gtt	gat	agc	gaa	acc	gtg	gcc	483
Arg	Ala	Gln	Glu	Gly	Gly	Ile	Ser	Glu	Val	Asp	Ser	Glu	Thr	Val	Ala	
			140					145						150		
tat	cat	ttc	cat	gaa	ccg	tta	ggc	gtg	gtg	ggg	cag	att	atc	ccg	tgg	531
Tyr	His	Phe	His	Glu	Pro	Leu	Gly	Val	Val	Gly	Gln	Ile	Ile	Pro	Trp	
155						160				165					170	
aac	ttc	ccg	ctg	ctg	atg	gcg	agc	tgg	aaa	atg	gct	ccc	gcg	ctg	gcg	579
Asn	Phe	Pro	Leu	Leu	Met	Ala	Ser	Trp	Lys	Met	Ala	Pro	Ala	Leu	Ala	
					175					180					185	
gcg	ggc	aac	tgt	gtg	gtg	ctg	aaa	ccc	gca	cgt	ctt	acc	ccg	ctt	tct	627
Ala	Gly	Asn	Cys	Val	Val	Leu	Lys	Pro	Ala	Arg	Leu	Thr	Pro	Leu	Ser	
			190					195						200		
gta	ctg	ctg	cta	atg	gaa	att	gtc	ggc	gat	tta	ctg	ccg	ccg	ggc	gtg	675
Val	Leu	Leu	Leu	Met	Glu	Ile	Val	Gly	Asp	Leu	Leu	Pro	Pro	Gly	Val	
			205					210						215		
gtg	aac	gtg	gtc	aac	ggc	gca	ggc	ggg	gta	att	ggc	gaa	tat	ctg	gcg	723
Val	Asn	Val	Val	Asn	Gly	Ala	Gly	Gly	Val	Ile	Gly	Glu	Tyr	Leu	Ala	
			220					225						230		
acc	tcg	aaa	cgc	atc	gcc	aaa	gtg	gcg	ttt	acc	ggc	tca	acg	gaa	gtg	771
Thr	Ser	Lys	Arg	Ile	Ala	Lys	Val	Ala	Phe	Thr	Gly	Ser	Thr	Glu	Val	
235						240				245					250	
ggc	caa	caa	att	atg	caa	tac	gca	acg	caa	aac	att	att	ccg	gtg	acg	819

Gly Gln Gln Ile Met Gln Tyr Ala Thr Gln Asn Ile Ile Pro Val Thr	
255	260 265
ctg gag ttg ggc ggt aag tcg cca aat atc gtc ttt gct gat gtg atg	867
Leu Glu Leu Gly Gly Lys Ser Pro Asn Ile Val Phe Ala Asp Val Met	
270	275 280
gat gaa gaa gat gcc ttt ttc gat aaa gcg ctg gaa ggc ttt gca ctg	915
Asp Glu Glu Asp Ala Phe Phe Asp Lys Ala Leu Glu Gly Phe Ala Leu	
285	290 295
ttt gcc ttt aac cag ggc gaa gtt tgc acc tgt ccg agt cgt gct tta	963
Phe Ala Phe Asn Gln Gly Glu Val Cys Thr Cys Pro Ser Arg Ala Leu	
300	305 310
gtg cag gaa tct atc tac gaa cgc ttt atg gaa cgc gcc atc cgc cgt	1011
Val Gln Glu Ser Ile Tyr Glu Arg Phe Met Glu Arg Ala Ile Arg Arg	
315	320 325 330
gtc gaa agc att cgt agc ggt aac ccg ctc gac agc gtg acg caa atg	1059
Val Glu Ser Ile Arg Ser Gly Asn Pro Leu Asp Ser Val Thr Gln Met	
335	340 345
ggc gcg cag gtt tct cac ggg caa ctg gaa acc atc ctc aac tac att	1107
Gly Ala Gln Val Ser His Gly Gln Leu Glu Thr Ile Leu Asn Tyr Ile	
350	355 360
gat atc ggt aaa aaa gag ggc gct gac gtg ctc aca ggc ggg cgg cgc	1155
Asp Ile Gly Lys Lys Glu Gly Ala Asp Val Leu Thr Gly Gly Arg Arg	
365	370 375
aag ctg ctg gaa ggt gaa ctg aaa gac ggc tac tac ctc gaa ccg acg	1203
Lys Leu Leu Glu Gly Glu Leu Lys Asp Gly Tyr Tyr Leu Glu Pro Thr	
380	385 390
att ctg ttt ggt cag aac aat atg cgg gtg ttc cag gag gag att ttt	1251
Ile Leu Phe Gly Gln Asn Asn Met Arg Val Phe Gln Glu Glu Ile Phe	
395	400 405 410
ggc ccg gtg ctg gcg gtg acc acc ttc aaa acg atg gaa gaa gcg ctg	1299
Gly Pro Val Leu Ala Val Thr Thr Phe Lys Thr Met Glu Glu Ala Leu	
415	420 425
gag ctg gcg aac gat acg caa tat ggc ctg ggc gcg ggc gtc tgg agc	1347
Glu Leu Ala Asn Asp Thr Gln Tyr Gly Leu Gly Ala Gly Val Trp Ser	
430	435 440
cgc aac ggt aat ctg gcc tat aag atg ggg cgc ggc ata cag gct ggg	1395

Arg Asn Gly Asn Leu Ala Tyr Lys Met Gly Arg Gly Ile Gln Ala Gly
 445 450 455

cgc gtg tgg acc aac tgt tat cac gct tac ccg gca cat gcg gcg ttt 1443
 Arg Val Trp Thr Asn Cys Tyr His Ala Tyr Pro Ala His Ala Ala Phe
 460 465 470

ggt ggc tac aaa caa tca ggt atc ggt cgc gaa acc cac aag atg atg 1491
 Gly Gly Tyr Lys Gln Ser Gly Ile Gly Arg Glu Thr His Lys Met Met
 475 480 485 490

ctg gag cat tac cag caa acc aag tgc ctg ctg gtg agc tac tcg gat 1539
 Leu Glu His Tyr Gln Gln Thr Lys Cys Leu Leu Val Ser Tyr Ser Asp
 495 500 505

aaa ccg ttg ggg ctg ttc taagagctcg aattcgc 1574
 Lys Pro Leu Gly Leu Phe
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<210> 8

<211> 512

<212> PRT

<213> Escherichia coli

<400> 8

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Trp Val Ala Pro Ala Asp Gly Glu Tyr Tyr Gln Asn Leu Thr Pro Val
 35 40 45

Thr Gly Gln Leu Leu Cys Glu Val Ala Ser Ser Gly Lys Arg Asp Ile
 50 55 60

Asp Leu Ala Leu Asp Ala Ala His Lys Val Lys Asp Lys Trp Ala His
 65 70 75 80

Thr Ser Val Gln Asp Arg Ala Ala Ile Leu Phe Lys Ile Ala Asp Arg
 85 90 95

Met Glu Gln Asn Leu Glu Leu Leu Ala Thr Ala Glu Thr Trp Asp Asn
 100 105 110

Gly Lys Pro Ile Arg Glu Thr Ser Ala Ala Asp Val Pro Leu Ala Ile

115		120		125
Asp His Phe Arg Tyr Phe Ala Ser Cys Ile Arg Ala Gln Glu Gly Gly				
130		135		140
Ile Ser Glu Val Asp Ser Glu Thr Val Ala Tyr His Phe His Glu Pro				
145		150		155
				160
Leu Gly Val Val Gly Gln Ile Ile Pro Trp Asn Phe Pro Leu Leu Met				
		165		170
				175
Ala Ser Trp Lys Met Ala Pro Ala Leu Ala Ala Gly Asn Cys Val Val				
		180		185
				190
Leu Lys Pro Ala Arg Leu Thr Pro Leu Ser Val Leu Leu Leu Met Glu				
		195		200
				205
Ile Val Gly Asp Leu Leu Pro Pro Gly Val Val Asn Val Val Asn Gly				
		210		215
				220
Ala Gly Gly Val Ile Gly Glu Tyr Leu Ala Thr Ser Lys Arg Ile Ala				
225		230		235
				240
Lys Val Ala Phe Thr Gly Ser Thr Glu Val Gly Gln Gln Ile Met Gln				
		245		250
				255
Tyr Ala Thr Gln Asn Ile Ile Pro Val Thr Leu Glu Leu Gly Gly Lys				
		260		265
				270
Ser Pro Asn Ile Val Phe Ala Asp Val Met Asp Glu Glu Asp Ala Phe				
		275		280
				285
Phe Asp Lys Ala Leu Glu Gly Phe Ala Leu Phe Ala Phe Asn Gln Gly				
		290		295
				300
Glu Val Cys Thr Cys Pro Ser Arg Ala Leu Val Gln Glu Ser Ile Tyr				
305		310		315
				320
Glu Arg Phe Met Glu Arg Ala Ile Arg Arg Val Glu Ser Ile Arg Ser				
		325		330
				335
Gly Asn Pro Leu Asp Ser Val Thr Gln Met Gly Ala Gln Val Ser His				
		340		345
				350
Gly Gln Leu Glu Thr Ile Leu Asn Tyr Ile Asp Ile Gly Lys Lys Glu				
		355		360
				365
Gly Ala Asp Val Leu Thr Gly Gly Arg Arg Lys Leu Leu Glu Gly Glu				

370		375		380
Leu Lys Asp Gly Tyr Tyr Leu Glu Pro Thr Ile Leu Phe Gly Gln Asn				
385		390		395
				400
Asn Met Arg Val Phe Gln Glu Glu Ile Phe Gly Pro Val Leu Ala Val				
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Thr Leu Ala Ala Leu Glu Gln Ala Leu Ala Lys Thr Pro Trp Ser Met
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Ser Asp Val Ser Arg Ile Tyr Leu Asn Glu Ala Ala Pro Val Ile Gly
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Asp Val Ala Met Glu Thr Ile Thr Glu Thr Ile Ile Thr Glu Ser Thr
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Met Ile Gly His Asn Pro Gln Thr Pro Gly Gly Val Gly Val Gly Val
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Gly Thr Thr Ile Ala Leu Gly Arg Leu Ala Thr Leu Pro Ala Ala Gln
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Tyr Ala Glu Gly Trp Ile Val Leu Ile Asp Asp Ala Val Asp Phe Leu
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Asp Ala Val Trp Trp Leu Asn Glu Ala Leu Asp Arg Gly Ile Asn Val
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Val Ala Ala Ile Leu Lys Lys Asp Asp Gly Val Leu Val Asn Asn Arg
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Leu Arg Lys Thr Leu Pro Val Val Asp Glu Val Thr Leu Leu Glu Gln
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Val Pro Glu Gly Val Met Ala Ala Val Glu Val Ala Ala Pro Gly Gln
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Val Val Arg Ile Leu Ser Asn Pro Tyr Gly Ile Ala Thr Phe Phe Gly
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Leu Ser Pro Glu Glu Thr Gln Ala Ile Val Pro Ile Ala Arg Ala Leu
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Ile Gly Asn Arg Ser Ala Val Val Leu Lys Thr Pro Gln Gly Asp Val
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Gln Ser Arg Val Ile Pro Ala Gly Asn Leu Tyr Ile Ser Gly Glu Lys
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Thr His Ala Gly Gly Met Leu Glu Arg Val Arg Lys Val Met Ala Ser
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Val Asp Thr Phe Ile Pro Arg Lys Val Gln Gly Gly Met Ala Gly Glu
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Cys Ala Met Glu Asn Ala Val Gly Met Ala Ala Met Val Lys Ala Asp
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Ile Ser Arg Gln Thr Leu Glu Tyr Gln Ala Gln Ile Ala Glu Gln Met
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Gln Arg His Ala Val Ala Arg Asn Phe Arg Arg Ala Ala Glu Leu Ile
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Ala Ile Pro Asp Glu Arg Ile Leu Ala Ile Tyr Asn Ala Leu Arg Pro
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Phe Arg Ser Ser Gln Ala Glu Leu Leu Ala Ile Ala Asp Glu Leu Glu
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Ile Gly Ile Gln Ser Lys Gly Thr Thr Val Ile His Gln Arg Asp Leu
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Leu Pro Leu Ser Asn Leu Glu Leu Phe Ser Gln Ala Pro Leu Leu Thr
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Leu Glu Thr Tyr Arg Gln Ile Gly Lys Asn Ala Ala Arg Tyr Ala Arg
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Lys Glu Ser Pro Ser Pro Val Pro Val Val Asn Asp Gln Met Val Arg
 100 105 110

Pro Lys Phe Met Ala Lys Ala Ala Leu Phe His Ile Lys Glu Thr Lys
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Ile Val Glu Leu Asp Gly Lys Arg Arg Asp Gln Phe Asp Met Ile Asp
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Arg Phe Ile Ala Asp Tyr Ala Ile Asn Val Glu Arg Thr Glu Gln Ala
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Met Arg Leu Glu Ala Val Glu Ile Ala Arg Met Leu Val Asp Ile His
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Val Ser Arg Glu Glu Ile Ile Ala Ile Thr Thr Ala Ile Thr Pro Ala
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Lys Ala Val Glu Val Met Ala Gln Met Asn Val Val Glu Met Met Met
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Ala Leu Gln Lys Met Arg Ala Arg Arg Thr Pro Ser Asn Gln Cys His
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Cameron, Douglas C.

<120> Production of 3-Hydroxypropionic Acid in Recombinant Organisms

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25 tac gac aaa ttc att gaa gag ttc aaa gcc gct tct gaa tcc atc aag 1011
 Tyr Asp Lys Phe Ile Glu Glu Phe Lys Ala Ala Ser Glu Ser Ile Lys
 315 320 325

gtg ggc gac cca ttc gat gaa tct act ttc caa ggt gca caa acc tct 1059
Val Gly Asp Pro Phe Asp Glu Ser Thr Phe Gln Gly Ala Gln Thr Ser
330 335 340 345

caa atg caa cta aac aaa atc ttg aaa tac gtt gac att ggt aag aat 1107
5 Gln Met Gln Leu Asn Lys Ile Leu Lys Tyr Val Asp Ile Gly Lys Asn
350 355 360

gaa ggt gct act ttg att acc ggt ggt gaa aga tta ggt agc aag ggt 1155
Glu Gly Ala Thr Leu Ile Thr Gly Gly Glu Arg Leu Gly Ser Lys Gly
365 370 375

10 tac ttc att aag cca act gtc ttt ggt gac gtt aag gaa gac atg aga 1203
Tyr Phe Ile Lys Pro Thr Val Phe Gly Asp Val Lys Glu Asp Met Arg
380 385 390

att gtc aaa gag gaa atc ttt ggc cct gtt gtc act gta acc aaa ttc 1251
Ile Val Lys Glu Glu Ile Phe Gly Pro Val Val Thr Val Thr Lys Phe
15 395 400 405

aaa tct gcc gac gaa gtc att aac atg gcg aac gat tct gaa tac ggg 1299
Lys Ser Ala Asp Glu Val Ile Asn Met Ala Asn Asp Ser Glu Tyr Gly
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20 Leu Ala Ala Gly Ile His Thr Ser Asn Ile Asn Thr Ala Leu Lys Val
430 435 440

gct gat aga gtt aat gcg ggt acg gtc tgg ata aac act tat aac gat 1395
Ala Asp Arg Val Asn Ala Gly Thr Val Trp Ile Asn Thr Tyr Asn Asp
445 450 455

25 ttc cac cac gca gtt cct ttc ggt ggg ttc aat gca tct ggt ttg ggc 1443
Phe His His Ala Val Pro Phe Gly Gly Phe Asn Ala Ser Gly Leu Gly
460 465 470

agg gaa atg tct gtt gat gct tta caa aac tac ttg caa gtt aaa gcg 1491
Arg Glu Met Ser Val Asp Ala Leu Gln Asn Tyr Leu Gln Val Lys Ala
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Ser Lys Gln Asn Lys Thr Phe Glu Val Ile Asn Pro Ser Thr Glu Glu
35 40 45
Glu Ile Cys His Ile Tyr Glu Gly Arg Glu Asp Asp Val Glu Glu Ala
50 55 60
20 Val Gln Ala Ala Asp Arg Ala Phe Ser Asn Gly Ser Trp Asn Gly Ile
65 70 75 80
Asp Pro Ile Asp Arg Gly Lys Ala Leu Tyr Arg Leu Ala Glu Leu Ile
85 90 95
Glu Gln Asp Lys Asp Val Ile Ala Ser Ile Glu Thr Leu Asp Asn Gly
25 100 105 110
Lys Ala Ile Ser Ser Ser Arg Gly Asp Val Asp Leu Val Ile Asn Tyr
115 120 125

Leu Lys Ser Ser Ala Gly Phe Ala Asp Lys Ile Asp Gly Arg Met Ile
 130 135 140

Asp Thr Gly Arg Thr His Phe Ser Tyr Thr Lys Arg Gln Pro Leu Gly
 145 150 155 160

5 Val Cys Gly Gln Ile Ile Pro Trp Asn Phe Pro Leu Leu Met Trp Ala
 165 170 175

Trp Lys Ile Ala Pro Ala Leu Val Thr Gly Asn Thr Val Val Leu Lys
 180 185 190

10 Thr Ala Glu Ser Thr Pro Leu Ser Ala Leu Tyr Val Ser Lys Tyr Ile
 195 200 205

Pro Gln Ala Gly Ile Pro Pro Gly Val Ile Asn Ile Val Ser Gly Phe
 210 215 220

Gly Lys Ile Val Val Glu Ala Ile Thr Asn His Pro Lys Ile Lys Lys
 225 230 235 240

15 Val Ala Phe Thr Gly Ser Thr Ala Thr Gly Arg His Ile Tyr Gln Ser
 245 250 255

Ala Ala Ala Gly Leu Lys Lys Val Thr Leu Glu Leu Gly Gly Lys Ser
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20 Pro Asn Ile Val Phe Ala Asp Ala Glu Leu Lys Lys Ala Val Gln Asn
 275 280 285

Ile Ile Leu Gly Ile Tyr Tyr Asn Ser Gly Glu Val Cys Cys Ala Gly
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Ser Arg Val Tyr Val Glu Glu Ser Ile Tyr Asp Lys Phe Ile Glu Glu
 305 310 315 320

25 Phe Lys Ala Ala Ser Glu Ser Ile Lys Val Gly Asp Pro Phe Asp Glu
 325 330 335

Ser Thr Phe Gln Gly Ala Gln Thr Ser Gln Met Gln Leu Asn Lys Ile
340 345 350

Leu Lys Tyr Val Asp Ile Gly Lys Asn Glu Gly Ala Thr Leu Ile Thr
355 360 365

5 Gly Gly Glu Arg Leu Gly Ser Lys Gly Tyr Phe Ile Lys Pro Thr Val
370 375 380

Phe Gly Asp Val Lys Glu Asp Met Arg Ile Val Lys Glu Glu Ile Phe
385 390 395 400

10 Gly Pro Val Val Thr Val Thr Lys Phe Lys Ser Ala Asp Glu Val Ile
405 410 415

Asn Met Ala Asn Asp Ser Glu Tyr Gly Leu Ala Ala Gly Ile His Thr
420 425 430

Ser Asn Ile Asn Thr Ala Leu Lys Val Ala Asp Arg Val Asn Ala Gly
435 440 445

15 Thr Val Trp Ile Asn Thr Tyr Asn Asp Phe His His Ala Val Pro Phe
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Gly Gly Phe Asn Ala Ser Gly Leu Gly Arg Glu Met Ser Val Asp Ala
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485 490 495

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      Ala Pro Asn Gln Gln Pro Glu Val Phe Cys Asn Gln Ile Phe Ile Asn
10              15              20              25

      aat gaa tgg cac gat gcc gtc agc agg aaa aca ttc ccc acc gtc aat      147
      Asn Glu Trp His Asp Ala Val Ser Arg Lys Thr Phe Pro Thr Val Asn
              30              35              40

      ccg tcc act gga gag gtc atc tgt cag gta gct gaa ggg gac aag gaa      195
15  Pro Ser Thr Gly Glu Val Ile Cys Gln Val Ala Glu Gly Asp Lys Glu
              45              50              55

      gat gtg gac aag gca cgt gaa ggc cgc ccg ggc gcc ttc cag ctg ggc      243
      Asp Val Asp Lys Ala Arg Glu Gly Arg Pro Gly Ala Phe Gln Leu Gly
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20  tca cct tgg cgc cgc atg gac gca tca cac agc ggc cgg ctg ctg aac      291
      Ser Pro Trp Arg Arg Met Asp Ala Ser His Ser Gly Arg Leu Leu Asn
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      cgc ctg gcc gat ctg atc gag cgg gac cgg acc tac ctg gcg gcc ttg      339
      Arg Leu Ala Asp Leu Ile Glu Arg Asp Arg Thr Tyr Leu Ala Ala Leu
25              95              100              105

      gag acc ctg gac aat ggc aag ccc tat gtc atc tcc tac ctg gtg gat      387
      Glu Thr Leu Asp Asn Gly Lys Pro Tyr Val Ile Ser Tyr Leu Val Asp
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5	Lys Tyr His Gly Lys Thr Ile Pro Ile Asp Gly Asp Phe Phe Ser Tyr	
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	aca cgc cat gaa cct gtg ggg gtg tgc ggg cag atc att ccg tgg aat	531
	Thr Arg His Glu Pro Val Gly Val Cys Gly Gln Ile Ile Pro Trp Asn	
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10	ttc ccg ctc ctg atg caa gca tgg aag ctg ggc cca gcc ttg gca act	579
	Phe Pro Leu Leu Met Gln Ala Trp Lys Leu Gly Pro Ala Leu Ala Thr	
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	Gly Asn Val Val Val Met Lys Val Ala Glu Gln Thr Pro Leu Thr Ala	
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	Leu Tyr Val Ala Asn Leu Ile Lys Glu Ala Gly Phe Pro Pro Gly Val	
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	220 225 230	
	tcc cat gag gat gtg gac aaa gtg gca ttc aca ggc tcc act gag att	771
	Ser His Glu Asp Val Asp Lys Val Ala Phe Thr Gly Ser Thr Glu Ile	
	235 240 245 250	
25	ggc cgc gta atc cag gtt gct gct ggg agc agc aac ctc aag aga gtg	819
	Gly Arg Val Ile Gln Val Ala Ala Gly Ser Ser Asn Leu Lys Arg Val	
	255 260 265	

acc ttg gag ctg ggg ggg aag agc ccc aac atc atc atg tca gat gcc 867
Thr Leu Glu Leu Gly Gly Lys Ser Pro Asn Ile Ile Met Ser Asp Ala
270 275 280

gat atg gat tgg gcc gtg gaa cag gcc cac ttc gcc ctg ttc ttc aac 915
5 Asp Met Asp Trp Ala Val Glu Gln Ala His Phe Ala Leu Phe Phe Asn
285 290 295

cag ggc cag tgc tgc tgt gcc ggc tcc cgg acc ttc gtg cag gag gac 963
Gln Gly Gln Cys Cys Cys Ala Gly Ser Arg Thr Phe Val Gln Glu Asp
300 305 310

10 atc tat gat gag ttt gtg gtg cgg agc gtt gcc cgg gcc aag tct cgg 1011
Ile Tyr Asp Glu Phe Val Val Arg Ser Val Ala Arg Ala Lys Ser Arg
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gtg gtc ggg aac ccc ttt gat agc aag acc gag cag ggg ccg cag gtg 1059
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15 335 340 345

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Asp Glu Thr Gln Phe Lys Lys Ile Leu Gly Tyr Ile Asn Thr Gly Lys
350 355 360

caa gag ggg gcg aag ctg ctg tgt ggt ggg ggc att gct gct gac cgt 1155
20 Gln Glu Gly Ala Lys Leu Leu Cys Gly Gly Gly Ile Ala Ala Asp Arg
365 370 375

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Gly Tyr Phe Ile Gln Pro Thr Val Phe Gly Asp Val Gln Asp Gly Met
380 385 390

25 acc atc gcc aag gag gag atc ttc ggg cca gtg atg cag atc ctg aag 1251
Thr Ile Ala Lys Glu Glu Ile Phe Gly Pro Val Met Gln Ile Leu Lys
395 400 405 410

ttc aag acc ata gag gag gtt gtt ggg aga gcc aac aat tcc acg tac 1299
Phe Lys Thr Ile Glu Glu Val Val Gly Arg Ala Asn Asn Ser Thr Tyr
415 420 425

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5 Gly Leu Ala Ala Ala Val Phe Thr Lys Asp Leu Asp Lys Ala Asn Tyr
430 435 440

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445 450 455

10 gtg ttt gga gcc cag tca ccc ttt ggt ggc tac aag atg tcg ggg agt 1443
Val Phe Gly Ala Gln Ser Pro Phe Gly Gly Tyr Lys Met Ser Gly Ser
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Gly Arg Glu Leu Gly Glu Tyr Gly Leu Gln Ala Tyr Thr Glu Val Lys
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<213> Homo sapiens

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Val Ser Arg Lys Thr Phe Pro Thr Val Asn Pro Ser Thr Gly Glu Val
35 40 45

Ile Cys Gln Val Ala Glu Gly Asp Lys Glu Asp Val Asp Lys Ala Arg
50 55 60

5 Glu Gly Arg Pro Gly Ala Phe Gln Leu Gly Ser Pro Trp Arg Arg Met
65 70 75 80

Asp Ala Ser His Ser Gly Arg Leu Leu Asn Arg Leu Ala Asp Leu Ile
85 90 95

10 Glu Arg Asp Arg Thr Tyr Leu Ala Ala Leu Glu Thr Leu Asp Asn Gly
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Lys Pro Tyr Val Ile Ser Tyr Leu Val Asp Leu Asp Met Val Leu Lys
115 120 125

Cys Leu Arg Tyr Tyr Ala Gly Trp Ala Asp Lys Tyr His Gly Lys Thr
130 135 140

15 Ile Pro Ile Asp Gly Asp Phe Phe Ser Tyr Thr Arg His Glu Pro Val
145 150 155 160

Gly Val Cys Gly Gln Ile Ile Pro Trp Asn Phe Pro Leu Leu Met Gln
165 170 175

20 Ala Trp Lys Leu Gly Pro Ala Leu Ala Thr Gly Asn Val Val Val Met
180 185 190

Lys Val Ala Glu Gln Thr Pro Leu Thr Ala Leu Tyr Val Ala Asn Leu
195 200 205

Ile Lys Glu Ala Gly Phe Pro Pro Gly Val Val Asn Ile Val Pro Gly
210 215 220

Phe Gly Pro Thr Ala Gly Ala Ala Ile Ala Ser His Glu Asp Val Asp
225 230 235 240

Lys Val Ala Phe Thr Gly Ser Thr Glu Ile Gly Arg Val Ile Gln Val
245 250 255

5 Ala Ala Gly Ser Ser Asn Leu Lys Arg Val Thr Leu Glu Leu Gly Gly
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Lys Ser Pro Asn Ile Ile Met Ser Asp Ala Asp Met Asp Trp Ala Val
275 280 285

10 Glu Gln Ala His Phe Ala Leu Phe Phe Asn Gln Gly Gln Cys Cys Cys
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Ala Gly Ser Arg Thr Phe Val Gln Glu Asp Ile Tyr Asp Glu Phe Val
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Val Arg Ser Val Ala Arg Ala Lys Ser Arg Val Val Gly Asn Pro Phe
325 330 335

15 Asp Ser Lys Thr Glu Gln Gly Pro Gln Val Asp Glu Thr Gln Phe Lys
340 345 350

Lys Ile Leu Gly Tyr Ile Asn Thr Gly Lys Gln Glu Gly Ala Lys Leu
355 360 365

20 Leu Cys Gly Gly Gly Ile Ala Ala Asp Arg Gly Tyr Phe Ile Gln Pro
370 375 380

Thr Val Phe Gly Asp Val Gln Asp Gly Met Thr Ile Ala Lys Glu Glu
385 390 395 400

Ile Phe Gly Pro Val Met Gln Ile Leu Lys Phe Lys Thr Ile Glu Glu
405 410 415

Val Val Gly Arg Ala Asn Asn Ser Thr Tyr Gly Leu Ala Ala Ala Val
420 425 430

Phe Thr Lys Asp Leu Asp Lys Ala Asn Tyr Leu Ser Gln Ala Leu Gln
435 440 445

5 Ala Gly Thr Val Trp Val Asn Cys Tyr Asp Val Phe Gly Ala Gln Ser
450 455 460

Pro Phe Gly Gly Tyr Lys Met Ser Gly Ser Gly Arg Glu Leu Gly Glu
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25 His Pro Met Tyr Ile Asp Gly Gln Phe Val Thr Trp Arg Gly Asp Ala
10 15 20

tgg att gat gtg gta aac cct gct aca gag gct gtc att tcc cgc ata 150
Trp Ile Asp Val Val Asn Pro Ala Thr Glu Ala Val Ile Ser Arg Ile
25 30 35

ccc gat ggt cag gcc gag gat gcc cgt aag gca atc gat gca gca gaa 198
5 Pro Asp Gly Gln Ala Glu Asp Ala Arg Lys Ala Ile Asp Ala Ala Glu
40 45 50

cgt gca caa cca gaa tgg gaa gcg ttg cct gct att gaa cgc gcc agt 246
Arg Ala Gln Pro Glu Trp Glu Ala Leu Pro Ala Ile Glu Arg Ala Ser
55 60 65 70

10 tgg ttg cgc aaa atc tcc gcc ggg atc cgc gaa cgc gcc agt gaa atc 294
Trp Leu Arg Lys Ile Ser Ala Gly Ile Arg Glu Arg Ala Ser Glu Ile
75 80 85

agt gcg ctg att gtt gaa gaa ggg ggc aag atc cag cag ctg gct gaa 342
Ser Ala Leu Ile Val Glu Glu Gly Gly Lys Ile Gln Gln Leu Ala Glu
15 90 95 100

gtc gaa gtg gct ttt act gcc gac tat atc gat tac atg gcg gag tgg 390
Val Glu Val Ala Phe Thr Ala Asp Tyr Ile Asp Tyr Met Ala Glu Trp
105 110 115

gca cgg cgt tac gag ggc gag att att caa agc gat cgt cca gga gaa 438
20 Ala Arg Arg Tyr Glu Gly Glu Ile Ile Gln Ser Asp Arg Pro Gly Glu
120 125 130

aat att ctt ttg ttt aaa cgt gcg ctt ggt gtg act acc ggc att ctg 486
Asn Ile Leu Leu Phe Lys Arg Ala Leu Gly Val Thr Thr Gly Ile Leu
135 140 145 150

25 ccg tgg aac ttc ccg ttc ttc ctc att gcc cgc aaa atg gct ccc gct 534
Pro Trp Asn Phe Pro Phe Phe Leu Ile Ala Arg Lys Met Ala Pro Ala
155 160 165

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	Leu Leu Thr Gly Asn Thr Ile Val Ile Lys Pro Ser Glu Phe Thr Thr	
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	aac aat gcg att gca ttc gcc aaa atc gtc gat gaa ata ggc ctt ccg	630
5	Asn Asn Ala Ile Ala Phe Ala Lys Ile Val Asp Glu Ile Gly Leu Pro	
	185 190 195	
	cgc gcc gtg ttt aac ctt gta ctg ggg cgt ggt gaa acc gtt ggg caa	678
	Arg Gly Val Phe Asn Leu Val Leu Gly Arg Gly Glu Thr Val Gly Gln	
	200 205 210	
10	gaa ctg gcg ggt aac cca aag gtc gca atg gtc agt atg aca ggc agc	726
	Glu Leu Ala Gly Asn Pro Lys Val Ala Met Val Ser Met Thr Gly Ser	
	215 220 225 230	
	gtc tct gca ggt gag aag atc atg gcg act gcg gcg aaa aac atc acc	774
	Val Ser Ala Gly Glu Lys Ile Met Ala Thr Ala Ala Lys Asn Ile Thr	
15	235 240 245	
	aaa gtg tgt ctg gaa ttg ggg ggt aaa gca cca gct atc gta atg gac	822
	Lys Val Cys Leu Glu Leu Gly Gly Lys Ala Pro Ala Ile Val Met Asp	
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20	Asp Ala Asp Leu Glu Leu Ala Val Lys Ala Ile Val Asp Ser Arg Val	
	265 270 275	
	att aat agt ggg caa gtg tgt aac tgt gca gaa cgt gtt tat gta cag	918
	Ile Asn Ser Gly Gln Val Cys Asn Cys Ala Glu Arg Val Tyr Val Gln	
	280 285 290	
25	aaa ggc att tat gat cag ttc gtc aat cgg ctg ggt gaa gcg atg cag	966
	Lys Gly Ile Tyr Asp Gln Phe Val Asn Arg Leu Gly Glu Ala Met Gln	
	295 300 305 310	

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315 320 325

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5 Pro Leu Ile Asn Ala Ala Ala Leu Glu Arg Val Glu Gln Lys Val Ala
330 335 340

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345 350 355

10 gag ggg aaa gga tat tat tat ccg ccg aca ttg ctg ctg gat gtt cgc 1158
Glu Gly Lys Gly Tyr Tyr Tyr Pro Pro Thr Leu Leu Leu Asp Val Arg
360 365 370

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Gln Glu Met Ser Ile Met His Glu Glu Thr Phe Gly Pro Val Leu Pro
15 375 380 385 390

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Val Val Ala Phe Asp Thr Leu Glu Asp Ala Ile Ser Met Ala Asn Asp
395 400 405

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410 415 420

gcg atg aaa gcc att aaa ggg ctg aag ttt ggt gaa act tac atc aac 1350
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425 430 435

25 cgt gaa aac ttc gaa gct atg caa ggc ttc cac gcc gga tgg cgt aaa 1398
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Ser Gly Ile Gly Gly Ala Asp Gly Lys His Gly Leu His Gly Tyr Leu
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35 40 45

Ala Ile Asp Ala Ala Glu Arg Ala Gln Pro Glu Trp Glu Ala Leu Pro
20 50 55 60

Ala Ile Glu Arg Ala Ser Trp Leu Arg Lys Ile Ser Ala Gly Ile Arg
65 70 75 80

Glu Arg Ala Ser Glu Ile Ser Ala Leu Ile Val Glu Glu Gly Gly Lys
85 90 95

25 Ile Gln Gln Leu Ala Glu Val Glu Val Ala Phe Thr Ala Asp Tyr Ile
100 105 110

Asp Tyr Met Ala Glu Trp Ala Arg Arg Tyr Glu Gly Glu Ile Ile Gln
115 120 125

Ser Asp Arg Pro Gly Glu Asn Ile Leu Leu Phe Lys Arg Ala Leu Gly
130 135 140

5 Val Thr Thr Gly Ile Leu Pro Trp Asn Phe Pro Phe Phe Leu Ile Ala
145 150 155 160

Arg Lys Met Ala Pro Ala Leu Leu Thr Gly Asn Thr Ile Val Ile Lys
165 170 175

10 Pro Ser Glu Phe Thr Thr Asn Asn Ala Ile Ala Phe Ala Lys Ile Val
180 185 190

Asp Glu Ile Gly Leu Pro Arg Gly Val Phe Asn Leu Val Leu Gly Arg
195 200 205

Gly Glu Thr Val Gly Gln Glu Leu Ala Gly Asn Pro Lys Val Ala Met
210 215 220

15 Val Ser Met Thr Gly Ser Val Ser Ala Gly Glu Lys Ile Met Ala Thr
225 230 235 240

Ala Ala Lys Asn Ile Thr Lys Val Cys Leu Glu Leu Gly Gly Lys Ala
245 250 255

20 Pro Ala Ile Val Met Asp Asp Ala Asp Leu Glu Leu Ala Val Lys Ala
260 265 270

Ile Val Asp Ser Arg Val Ile Asn Ser Gly Gln Val Cys Asn Cys Ala
275 280 285

Glu Arg Val Tyr Val Gln Lys Gly Ile Tyr Asp Gln Phe Val Asn Arg
290 295 300

Leu Gly Glu Ala Met Gln Ala Val Gln Phe Gly Asn Pro Ala Glu Arg
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Asn Asp Ile Ala Met Gly Pro Leu Ile Asn Ala Ala Ala Leu Glu Arg
325 330 335

5 Val Glu Gln Lys Val Ala Arg Ala Val Glu Glu Gly Ala Arg Val Ala
340 345 350

Phe Gly Gly Lys Ala Val Glu Gly Lys Gly Tyr Tyr Tyr Pro Pro Thr
355 360 365

10 Leu Leu Leu Asp Val Arg Gln Glu Met Ser Ile Met His Glu Glu Thr
370 375 380

Phe Gly Pro Val Leu Pro Val Val Ala Phe Asp Thr Leu Glu Asp Ala
385 390 395 400

Ile Ser Met Ala Asn Asp Ser Asp Tyr Gly Leu Thr Ser Ser Ile Tyr
405 410 415

15 Thr Gln Asn Leu Asn Val Ala Met Lys Ala Ile Lys Gly Leu Lys Phe
420 425 430

Gly Glu Thr Tyr Ile Asn Arg Glu Asn Phe Glu Ala Met Gln Gly Phe
435 440 445

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Lys Pro Gly Glu Tyr Gly Phe Pro Leu Lys Leu Lys Ala Arg Tyr Asp
           15             20             25

15 aac ttt att ggc ggc gaa tgg gta gcc cct gcc gac ggc gag tat tac   147
Asn Phe Ile Gly Gly Glu Trp Val Ala Pro Ala Asp Gly Glu Tyr Tyr
           30             35             40

cag aat ctg acg ccg gtg acc ggg cag ctg ctg tgc gaa gtg gcg tct   195
Gln Asn Leu Thr Pro Val Thr Gly Gln Leu Leu Cys Glu Val Ala Ser
20      45             50             55

tcg ggc aaa cga gac atc gat ctg gcg ctg gat gct gcg cac aaa gtg   243
Ser Gly Lys Arg Asp Ile Asp Leu Ala Leu Asp Ala Ala His Lys Val
           60             65             70

aaa gat aaa tgg gcg cac acc tcg gtg cag gat cgt gcg gcg att ctg   291
25 Lys Asp Lys Trp Ala His Thr Ser Val Gln Asp Arg Ala Ala Ile Leu
           75             80             85             90

ttt aag att gcc gat cga atg gaa caa aac ctc gag ctg tta gcg aca   339
Phe Lys Ile Ala Asp Arg Met Glu Gln Asn Leu Glu Leu Leu Ala Thr
           95             100            105

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gct gaa acc tgg gat aac ggc aaa ccc att cgc gaa acc agt gct gcg 387
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gat gta cgc ctg gcg att gac cat ttc cgc tat ttc gcc tcg tgt att 435
5 Asp Val Pro Leu Ala Ile Asp His Phe Arg Tyr Phe Ala Ser Cys Ile
125 130 135

cgg gcg cag gaa ggt ggg atc agt gaa gtt gat agc gaa acc gtg gcc 483
Arg Ala Gln Glu Gly Gly Ile Ser Glu Val Asp Ser Glu Thr Val Ala
140 145 150

10 tat cat ttc cat gaa cgc tta ggc gtg gtg ggg cag att atc ccg tgg 531
Tyr His Phe His Glu Pro Leu Gly Val Val Gly Gln Ile Ile Pro Trp
155 160 165 170

aac ttc ccg ctg ctg atg gcg agc tgg aaa atg gct ccc gcg ctg gcg 579
Asn Phe Pro Leu Leu Met Ala Ser Trp Lys Met Ala Pro Ala Leu Ala
15 175 180 185

gcg ggc aac tgt gtg gtg ctg aaa ccc gca cgt ctt acc ccg ctt tct 627
Ala Gly Asn Cys Val Val Leu Lys Pro Ala Arg Leu Thr Pro Leu Ser
190 195 200

gta ctg ctg cta atg gaa att gtc ggt gat tta ctg ccg ccg ggc gtg 675
20 Val Leu Leu Leu Met Glu Ile Val Gly Asp Leu Leu Pro Pro Gly Val
205 210 215

gtg aac gtg gtc aat ggc gca ggt ggg gta att ggc gaa tat ctg gcg 723
Val Asn Val Val Asn Gly Ala Gly Gly Val Ile Gly Glu Tyr Leu Ala
220 225 230

25 acc tcg aaa cgc atc gcc aaa gtg gcg ttt acc ggc tca acg gaa gtg 771
Thr Ser Lys Arg Ile Ala Lys Val Ala Phe Thr Gly Ser Thr Glu Val
235 240 245 250

ggc caa caa att atg caa tac gca acg caa aac att att ccg gtg acg 819
Gly Gln Gln Ile Met Gln Tyr Ala Thr Gln Asn Ile Ile Pro Val Thr
255 260 265

ctg gag ttg ggc ggt aag tcg cca aat atc gtc ttt gct gat gtg atg 867
5 Leu Glu Leu Gly Gly Lys Ser Pro Asn Ile Val Phe Ala Asp Val Met
270 275 280

gat gaa gaa gat gcc ttt ttc gat aaa gcg ctg gaa ggc ttt gca ctg 915
Asp Glu Glu Asp Ala Phe Phe Asp Lys Ala Leu Glu Gly Phe Ala Leu
285 290 295

10 ttt gcc ttt aac cag ggc gaa gtt tgc acc tgt ccg agt cgt gct tta 963
 Phe Ala Phe Asn Gln Gly Glu Val Cys Thr Cys Pro Ser Arg Ala Leu
 300 305 310

gtg cag gaa tct atc tac gaa cgc ttt atg gaa cgc gcc atc cgc cgt 1011
Val Gln Glu Ser Ile Tyr Glu Arg Phe Met Glu Arg Ala Ile Arg Arg
15 315 320 325 330

gtc gaa agc att cgt agc ggt aac cgg ctc gac agc gtg acg caa atg 1059
Val Glu Ser Ile Arg Ser Gly Asn Pro Leu Asp Ser Val Thr Gln Met
335 340 345

ggc gcg cag gtt tct cac ggg caa ctg gaa acc atc ctc aac tac att 1107
20 Gly Ala Gln Val Ser His Gly Gln Leu Glu Thr Ile Leu Asn Tyr Ile
 350 355 360

gat atc ggt aaa aaa gag ggc gct gac gtg ctc aca ggc ggg cgg cgc 1155
Asp Ile Gly Lys Lys Glu Gly Ala Asp Val Leu Thr Gly Gly Arg Arg
365 370 375

25 aag ctg ctg gaa ggt gaa ctg aaa gac ggc tac tac ctc gaa ccg acg 1203
Lys Leu Leu Glu Gly Glu Leu Lys Asp Gly Tyr Tyr Leu Glu Pro Thr
380 385 390

att ctg ttt ggt cag aac aat atg cgg gtg ttc cag gag gag att ttt 1251
Ile Leu Phe Gly Gln Asn Asn Met Arg Val Phe Gln Glu Glu Ile Phe
395 400 405 410

ggc ccg gtg ctg gcg gtg acc acc ttc aaa acg atg gaa gaa gcg ctg 1299
5 Gly Pro Val Leu Ala Val Thr Thr Phe Lys Thr Met Glu Glu Ala Leu
415 420 425

gag ctg gcg aac gat acg caa tat ggc ctg ggc gcg ggc gtc tgg agc 1347
Glu Leu Ala Asn Asp Thr Gln Tyr Gly Leu Gly Ala Gly Val Trp Ser
430 435 440

10 cgc aac ggt aat ctg gcc tat aag atg ggg cgc ggc ata cag gct ggg 1395
Arg Asn Gly Asn Leu Ala Tyr Lys Met Gly Arg Gly Ile Gln Ala Gly
445 450 455

cgc gtg tgg acc aac tgt tat cac gct tac ccg gca cat gcg gcg ttt 1443
Arg Val Trp Thr Asn Cys Tyr His Ala Tyr Pro Ala His Ala Ala Phe
15 460 465 470

ggt ggc tac aaa caa tca ggt atc ggt cgc gaa acc cac aag atg atg 1491
Gly Gly Tyr Lys Gln Ser Gly Ile Gly Arg Glu Thr His Lys Met Met
475 480 485 490

ctg gag cat tac cag caa acc aag tgc ctg ctg gtg agc tac tcg gat 1539
20 Leu Glu His Tyr Gln Gln Thr Lys Cys Leu Leu Val Ser Tyr Ser Asp
495 500 505

aaa ccg ttg ggg ctg ttc taagagctcg aattcgc 1574
Lys Pro Leu Gly Leu Phe
510

<210> 8
<211> 512
<212> PRT
<213> Escherichia coli

5 <400> 8

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1 5 10 15

Phe Pro Leu Lys Leu Lys Ala Arg Tyr Asp Asn Phe Ile Gly Gly Glu
20 25 30

10 Trp Val Ala Pro Ala Asp Gly Glu Tyr Tyr Gln Asn Leu Thr Pro Val
35 40 45

Thr Gly Gln Leu Leu Cys Glu Val Ala Ser Ser Gly Lys Arg Asp Ile
50 55 60

Asp Leu Ala Leu Asp Ala Ala His Lys Val Lys Asp Lys Trp Ala His
15 65 70 75 80

Thr Ser Val Gln Asp Arg Ala Ala Ile Leu Phe Lys Ile Ala Asp Arg
85 90 95

Met Glu Gln Asn Leu Glu Leu Leu Ala Thr Ala Glu Thr Trp Asp Asn
100 105 110

20 Gly Lys Pro Ile Arg Glu Thr Ser Ala Ala Asp Val Pro Leu Ala Ile
115 120 125

Asp His Phe Arg Tyr Phe Ala Ser Cys Ile Arg Ala Gln Glu Gly Gly
130 135 140

Ile Ser Glu Val Asp Ser Glu Thr Val Ala Tyr His Phe His Glu Pro
25 145 150 155 160

Leu Gly Val Val Gly Gln Ile Ile Pro Trp Asn Phe Pro Leu Leu Met
165 170 175

Ala Ser Trp Lys Met Ala Pro Ala Leu Ala Ala Gly Asn Cys Val Val
180 185 190

5 Leu Lys Pro Ala Arg Leu Thr Pro Leu Ser Val Leu Leu Leu Met Glu
195 200 205

Ile Val Gly Asp Leu Leu Pro Pro Gly Val Val Asn Val Val Asn Gly
210 215 220

Ala Gly Gly Val Ile Gly Glu Tyr Leu Ala Thr Ser Lys Arg Ile Ala
10 225 230 235 240

Lys Val Ala Phe Thr Gly Ser Thr Glu Val Gly Gln Gln Ile Met Gln
245 250 255

Tyr Ala Thr Gln Asn Ile Ile Pro Val Thr Leu Glu Leu Gly Gly Lys
260 265 270

15 Ser Pro Asn Ile Val Phe Ala Asp Val Met Asp Glu Glu Asp Ala Phe
275 280 285

Phe Asp Lys Ala Leu Glu Gly Phe Ala Leu Phe Ala Phe Asn Gln Gly
290 295 300

Glu Val Cys Thr Cys Pro Ser Arg Ala Leu Val Gln Glu Ser Ile Tyr
20 305 310 315 320

Glu Arg Phe Met Glu Arg Ala Ile Arg Arg Val Glu Ser Ile Arg Ser
325 330 335

Gly Asn Pro Leu Asp Ser Val Thr Gln Met Gly Ala Gln Val Ser His
340 345 350

Gly Gln Leu Glu Thr Ile Leu Asn Tyr Ile Asp Ile Gly Lys Lys Glu
355 360 365

Gly Ala Asp Val Leu Thr Gly Gly Arg Arg Lys Leu Leu Glu Gly Glu
370 375 380

5 Leu Lys Asp Gly Tyr Tyr Leu Glu Pro Thr Ile Leu Phe Gly Gln Asn
385 390 395 400

Asn Met Arg Val Phe Gln Glu Glu Ile Phe Gly Pro Val Leu Ala Val
405 410 415

10 Thr Thr Phe Lys Thr Met Glu Glu Ala Leu Glu Leu Ala Asn Asp Thr
420 425 430

Gln Tyr Gly Leu Gly Ala Gly Val Trp Ser Arg Asn Gly Asn Leu Ala
435 440 445

Tyr Lys Met Gly Arg Gly Ile Gln Ala Gly Arg Val Trp Thr Asn Cys
450 455 460

15 Tyr His Ala Tyr Pro Ala His Ala Ala Phe Gly Gly Tyr Lys Gln Ser
465 470 475 480

Gly Ile Gly Arg Glu Thr His Lys Met Met Leu Glu His Tyr Gln Gln
485 490 495

20 Thr Lys Cys Leu Leu Val Ser Tyr Ser Asp Lys Pro Leu Gly Leu Phe
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<210> 9

<211> 5267

<212> DNA

<213> Klebsiella pneumoniae

<220>

<223> Location complement 300..2153

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<223> Locaton complement 2594..3034

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<223> Location complement 2191..4858

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<210> 10

<211> 607

30 <212> PRT

<213> *Klebsiella pneumoniae*

<400> 10

Met Pro Leu Ile Ala Gly Ile Asp Ile Gly Asn Ala Thr Thr Glu Val

1

5

10

15

Ala Leu Ala Ser Asp Tyr Pro Gln Ala Arg Ala Phe Val Ala Ser Gly
20 25 30

Ile Val Ala Thr Thr Gly Met Lys Gly Thr Arg Asp Asn Ile Ala Gly
35 40 45

5 Thr Leu Ala Ala Leu Glu Gln Ala Leu Ala Lys Thr Pro Trp Ser Met
50 55 60

Ser Asp Val Ser Arg Ile Tyr Leu Asn Glu Ala Ala Pro Val Ile Gly
65 70 75 80

10 Asp Val Ala Met Glu Thr Ile Thr Glu Thr Ile Ile Thr Glu Ser Thr
85 90 95

Met Ile Gly His Asn Pro Gln Thr Pro Gly Gly Val Gly Val Gly Val
100 105 110

Gly Thr Thr Ile Ala Leu Gly Arg Leu Ala Thr Leu Pro Ala Ala Gln
115 120 125

15 Tyr Ala Glu Gly Trp Ile Val Leu Ile Asp Asp Ala Val Asp Phe Leu
130 135 140

Asp Ala Val Trp Trp Leu Asn Glu Ala Leu Asp Arg Gly Ile Asn Val
145 150 155 160

20 Val Ala Ala Ile Leu Lys Lys Asp Asp Gly Val Leu Val Asn Asn Arg
165 170 175

Leu Arg Lys Thr Leu Pro Val Val Asp Glu Val Thr Leu Leu Glu Gln
180 185 190

Val Pro Glu Gly Val Met Ala Ala Val Glu Val Ala Ala Pro Gly Gln
195 200 205

Val Val Arg Ile Leu Ser Asn Pro Tyr Gly Ile Ala Thr Phe Phe Gly
210 215 220

Leu Ser Pro Glu Glu Thr Gln Ala Ile Val Pro Ile Ala Arg Ala Leu
225 230 235 240

5 Ile Gly Asn Arg Ser Ala Val Val Leu Lys Thr Pro Gln Gly Asp Val
245 250 255

Gln Ser Arg Val Ile Pro Ala Gly Asn Leu Tyr Ile Ser Gly Glu Lys
260 265 270

Arg Arg Gly Glu Ala Asp Val Ala Glu Gly Ala Glu Ala Ile Met Gln
10 275 280 285

Ala Met Ser Ala Cys Ala Pro Val Arg Asp Ile Arg Gly Glu Pro Gly
290 295 300

Thr His Ala Gly Gly Met Leu Glu Arg Val Arg Lys Val Met Ala Ser
305 310 315 320

15 Leu Thr Gly His Glu Met Ser Ala Ile Tyr Ile Gln Asp Leu Leu Ala
325 330 335

Val Asp Thr Phe Ile Pro Arg Lys Val Gln Gly Gly Met Ala Gly Glu
340 345 350

Cys Ala Met Glu Asn Ala Val Gly Met Ala Ala Met Val Lys Ala Asp
20 355 360 365

Arg Leu Gln Met Gln Val Ile Ala Arg Glu Leu Ser Ala Arg Leu Gln
370 375 380

Thr Glu Val Val Val Gly Gly Val Glu Ala Asn Met Ala Ile Ala Gly
385 390 395 400

Ala Leu Thr Thr Pro Gly Cys Ala Ala Pro Leu Ala Ile Leu Asp Leu
405 410 415

Gly Ala Gly Ser Thr Asp Ala Ala Ile Val Asn Ala Glu Gly Gln Ile
420 425 430

5 Thr Ala Val His Leu Ala Gly Ala Gly Asn Met Val Ser Leu Leu Ile
435 440 445

Lys Thr Glu Leu Gly Leu Glu Asp Leu Ser Leu Ala Glu Ala Ile Lys
450 455 460

10 Lys Tyr Pro Leu Ala Lys Val Glu Ser Leu Phe Ser Ile Arg His Glu
465 470 475 480

Asn Gly Ala Val Glu Phe Phe Arg Glu Ala Leu Ser Pro Ala Val Phe
485 490 495

Ala Lys Val Val Tyr Ile Lys Glu Gly Glu Leu Val Pro Ile Asp Asn
500 505 510

15 Ala Ser Pro Leu Glu Lys Ile Arg Leu Val Arg Arg Gln Ala Lys Glu
515 520 525

Lys Val Phe Val Thr Asn Cys Leu Arg Ala Leu Arg Gln Val Ser Pro
530 535 540

Gly Gly Ser Ile Arg Asp Ile Ala Phe Val Val Leu Val Gly Gly Ser
20 545 550 555 560

Ser Leu Asp Phe Glu Ile Pro Gln Leu Ile Thr Glu Ala Leu Ser His
565 570 575

Tyr Gly Val Val Ala Gly Gln Gly Asn Ile Arg Gly Thr Glu Gly Pro
580 585 590

Arg Asn Ala Val Ala Thr Gly Leu Leu Leu Ala Gly Gln Ala Asn
595 600 605

<210> 11

<211> 141

5 <212> PRT

<213> *Klebsiella pneumoniae*

<400> 11

Met Ser Glu Lys Thr Met Arg Val Gln Asp Tyr Pro Leu Ala Thr Arg
1 5 10 15

10 Cys Pro Glu His Ile Leu Thr Pro Thr Gly Lys Pro Leu Thr Asp Ile
20 25 30

Thr Leu Glu Lys Val Leu Ser Gly Glu Val Gly Pro Gln Asp Val Arg
35 40 45

Ile Ser Arg Gln Thr Leu Glu Tyr Gln Ala Gln Ile Ala Glu Gln Met
15 50 55 60

Gln Arg His Ala Val Ala Arg Asn Phe Arg Arg Ala Ala Glu Leu Ile
65 70 75 80

Ala Ile Pro Asp Glu Arg Ile Leu Ala Ile Tyr Asn Ala Leu Arg Pro
85 90 95

20 Phe Arg Ser Ser Gln Ala Glu Leu Leu Ala Ile Ala Asp Glu Leu Glu
100 105 110

His Thr Trp His Ala Thr Val Asn Ala Ala Phe Val Arg Glu Ser Ala
115 120 125

Glu Val Tyr Gln Gln Arg His Lys Leu Arg Lys Gly Ser
25 130 135 140

<210> 12
<211> 146
<212> PRT
<213> *Klebsiella pneumoniae*

5 <400> 12

Met Pro His Gly Ala Ile Leu Lys Glu Leu Ile Ala Gly Val Glu Glu
1 5 10 15

Glu Gly Leu His Ala Arg Val Val Arg Ile Leu Arg Thr Ser Asp Val
20 25 30

10 Ser Phe Met Ala Trp Asp Ala Ala Asn Leu Ser Gly Ser Gly Ile Gly
35 40 45

Ile Gly Ile Gln Ser Lys Gly Thr Thr Val Ile His Gln Arg Asp Leu
50 55 60

Leu Pro Leu Ser Asn Leu Glu Leu Phe Ser Gln Ala Pro Leu Leu Thr
15 65 70 75 80

Leu Glu Thr Tyr Arg Gln Ile Gly Lys Asn Ala Ala Arg Tyr Ala Arg
85 90 95

Lys Glu Ser Pro Ser Pro Val Pro Val Val Asn Asp Gln Met Val Arg
100 105 110

20 Pro Lys Phe Met Ala Lys Ala Ala Leu Phe His Ile Lys Glu Thr Lys
115 120 125

His Val Val Gln Asp Ala Glu Pro Val Thr Leu His Ile Asp Leu Val
130 135 140

Arg Glu
25 145

<210> 13
<211> 555
<212> PRT
<213> *Klebsiella pneumoniae*

5 <400> 13

Met Lys Arg Ser Lys Arg Phe Ala Val Leu Ala Gln Arg Pro Val Asn
1 5 10 15

Gln Asp Gly Leu Ile Gly Glu Trp Pro Glu Glu Gly Leu Ile Ala Met
20 25 30

10 Asp Ser Pro Phe Asp Pro Val Ser Ser Val Lys Val Asp Asn Gly Leu
35 40 45

Ile Val Glu Leu Asp Gly Lys Arg Arg Asp Gln Phe Asp Met Ile Asp
50 55 60

Arg Phe Ile Ala Asp Tyr Ala Ile Asn Val Glu Arg Thr Glu Gln Ala
15 65 70 75 80

Met Arg Leu Glu Ala Val Glu Ile Ala Arg Met Leu Val Asp Ile His
85 90 95

Val Ser Arg Glu Glu Ile Ile Ala Ile Thr Thr Ala Ile Thr Pro Ala
100 105 110

20 Lys Ala Val Glu Val Met Ala Gln Met Asn Val Val Glu Met Met Met
115 120 125

Ala Leu Gln Lys Met Arg Ala Arg Arg Thr Pro Ser Asn Gln Cys His
130 135 140

Val Thr Asn Leu Lys Asp Asn Pro Val Gln Ile Ala Ala Asp Ala Ala
25 145 150 155 160

Glu Ala Gly Ile Arg Gly Phe Ser Glu Gln Glu Thr Thr Val Gly Ile
165 170 175

Ala Arg Tyr Ala Pro Phe Asn Ala Leu Ala Leu Leu Val Gly Ser Gln
180 185 190

5 Cys Gly Arg Pro Gly Val Leu Thr Gln Cys Ser Val Glu Glu Ala Thr
195 200 205

Glu Leu Glu Leu Gly Met Arg Gly Leu Thr Ser Tyr Ala Glu Thr Val
210 215 220

Ser Val Tyr Gly Thr Glu Ala Val Phe Thr Asp Gly Asp Asp Thr Pro
10 225 230 235 240

Trp Ser Lys Ala Phe Leu Ala Ser Ala Tyr Ala Ser Arg Gly Leu Lys
245 250 255

Met Arg Tyr Thr Ser Gly Thr Gly Ser Glu Ala Leu Met Gly Tyr Ser
260 265 270

15 Glu Ser Lys Ser Met Leu Tyr Leu Glu Ser Arg Cys Ile Phe Ile Thr
275 280 285

Lys Gly Ala Gly Val Gln Gly Leu Gln Asn Gly Ala Val Ser Cys Ile
290 295 300

Gly Met Thr Gly Ala Val Pro Ser Gly Ile Arg Ala Val Leu Ala Glu
20 305 310 315 320

Asn Leu Ile Ala Ser Met Leu Asp Leu Glu Val Ala Ser Ala Asn Asp
325 330 335

Gln Thr Phe Ser His Ser Asp Ile Arg Arg Thr Ala Arg Thr Leu Met
340 345 350

Gln Met Leu Pro Gly Thr Asp Phe Ile Phe Ser Gly Tyr Ser Ala Val
355 360 365

Pro Asn Tyr Asp Asn Met Phe Ala Gly Ser Asn Phe Asp Ala Glu Asp
370 375 380

5 Phe Asp Asp Tyr Asn Ile Leu Gln Arg Asp Leu Met Val Asp Gly Gly
385 390 395 400

Leu Arg Pro Val Thr Glu Ala Glu Thr Ile Ala Ile Arg Gln Lys Ala
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10 Ala Arg Ala Ile Gln Ala Val Phe Arg Glu Leu Gly Leu Pro Pro Ile
420 425 430

Ala Asp Glu Glu Val Glu Ala Ala Thr Tyr Ala His Gly Ser Asn Glu
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Met Pro Pro Arg Asn Val Val Glu Asp Leu Ser Ala Val Glu Glu Met
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15 Met Lys Arg Asn Ile Thr Gly Leu Asp Ile Val Gly Ala Leu Ser Arg
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Ser Gly Phe Glu Asp Ile Ala Ser Asn Ile Leu Asn Met Leu Arg Gln
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20 Arg Val Thr Gly Asp Tyr Leu Gln Thr Ser Ala Ile Leu Asp Arg Gln
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Phe Glu Val Val Ser Ala Val Asn Asp Ile Asn Asp Tyr Gln Gly Pro
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34

15 QBMAD\223318

CLAIM OR CLAIMS

I/WE CLAIM:

1. A method for producing 3-hydroxypropionic acid comprising the steps of
providing in a fermenter a recombinant microorganism which expresses genes
5 for non-native enzymes which are capable of catalyzing the production of 3-
hydroxypropionic acid from glycerol;
providing a source of glycerol or glucose for the recombinant microorganism,
and
fermenting the microorganism under conditions which result in the accumulation
10 of 3-hydroxypropionic acid.
2. A method for producing 3-hydroxypropionic acid comprising the steps of
providing in a fermenter a recombinant microorganism which carries genetic
constructions for the expression of a glycerol dehydratase and an aldehyde
dehydrogenase which are capable of catalyzing the production of 3-hydroxypropionic
15 acid from glycerol;
providing a source of glycerol or glucose for the recombinant microorganism,
and
fermenting the microorganism under conditions which result in the accumulation
of 3-hydroxypropionic acid.

3. A method for producing 3-hydroxypropionic acid comprising the steps of providing in a fermenter a recombinant microorganism which carries a genetic construct which expresses the *dhaB* gene from *Klebsiella pneumoniae* and a gene for an aldehyde dehydrogenase, which are capable of catalyzing the production of 3-hydroxypropionic acid from glycerol;
- 5 providing a source of glycerol or glucose for the recombinant microorganism, and
- fermenting the microorganism under conditions which result in the accumulation of 3-hydroxypropionic acid.

- 10 4. The method of claim 3 wherein the gene for the aldehyde dehydrogenase is selected from the group consisting of *ALDH4*, *ALD2*, *aldA* and *aldB*.

5. The method of claim 3 wherein the aldehyde dehydrogenase is selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6 and SEQ ID NO:8.

6. A recombinant *E. coli* host comprising in its inheritable genetic materials
- 15 foreign genes encoding a glycerol dehydratase and an aldehyde dehydrogenase, such that the host is capable of producing 3-hydroxypropionic acid from glycerol.

7. A recombinant *E. coli* host comprising in its inheritable genetic materials the *dhaB* gene from *Klebsiella pneumoniae* and the *ald4* gene from *Saccharomyces cerevisiae*, such that the host is capable of producing 3-hydroxypropionic from glycerol.

8. A bacterial host comprising in its inheritable genetic material a genetic construction encoding for the expression of a glycerol dehydratase enzyme and an aldehyde dehydrogenase enzyme, such that the bacterial host is capable of converting glycerol to 3-hydroxypropionic acid.

5 9. The bacterial host of claim 8 wherein the glycerol dehydratase from *Klebsiella pneumoniae*.

10 10. The bacterial host of claim 8 wherein the gene encoding the glycerol dehydratase is the *dhaB* gene from *Klebsiella pneumoniae*.

10 11. The bacterial host of claim 8 wherein the aldehyde dehydrogenase is selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6 and SEQ ID NO:8.

 12. The bacterial host of claim 8 wherein the gene for the aldehyde dehydrogenase is selected from the group consisting of *ALDH4*, *ALD2*, *aldA* and *aldB*.

ABSTRACT OF THE DISCLOSURE

The production of 3-hydroxypropionic acid (3-HP) from glycerol in a bacterial host is described. 3-HP is a useful feedstock for the production of polymeric materials. The genetic engineering of a bacterial host with two enzymes is sufficient to enable
5 production of 3-HP. One enzyme is a glycerol dehydratase and the other is an aldehyde dehydrogenase.

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0010/PTO Rev. 6/95 U.S. Department of Commerce Patent and Trademark Office DECLARATION FOR UTILITY OR DESIGN PATENT APPLICATION <input type="checkbox"/> Declaration Submitted with Initial Filing OR <input checked="" type="checkbox"/> Declaration Submitted after Initial Filing	Attorney Docket Number	960296.96617
	First Named Inventor	Patrick F. Suthers
	COMPLETE IF KNOWN	
	Application Number	09/830,751
	Filing Date	08/30/1999
	Group Art Unit	
	Examiner Name	

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe that I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

PRODUCTION OF 3-HYDROXYPROPIONIC ACID IN RECOMBINANT ORGANISMS

the specification of which (Title of the Invention)

☐ is attached hereto

OR

☒ was filed on (MM/DD/YYYY) 08/30/2000 as United States Application Number or PCT International

Application Number PCT/US00/23878 and was amended on (MM/DD/YYYY) (if applicable).

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations § 1.56.

I hereby claim foreign priority benefits under Title 35, United States Code § 119(a)-(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate or § 365(a) of any PCT international application which designated at least one country other than the United States of America, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or any PCT international application having a filing date before that of the application on which priority is claimed.

Prior Foreign Application Number(s)	Country	Foreign Filing Date (MM/DD/YYYY)	Priority Not Claimed	Certified Copy Attached? YES	NO
			<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
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			<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
			<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

☐ Additional foreign applications numbers are listed on a supplemental priority sheet attached hereto:

I hereby claim the benefit under Title 35, United States Code § 119(e) of any United States provisional application(s) listed below.

Application Number(s)	Filing Date (MM/DD/YYYY)	<input type="checkbox"/> Additional provisional application numbers are listed on a supplemental priority sheet attached hereto.
60/151,440	08/30/1999	

Burden Hour Statement: This form is estimated to take .4 hours to complete. Time will vary depending upon the needs of the individual case. Any comments on the amount of time you are required to complete this form should be sent to the Chief Information Officer, Patent and Trademark Office, Washington, DC 20231. DO NOT SEND FEES OR COMPLETED FORMS TO THIS ADDRESS. SEND TO: Commissioner for Patents, Washington, DC 20231. QBMA01337264

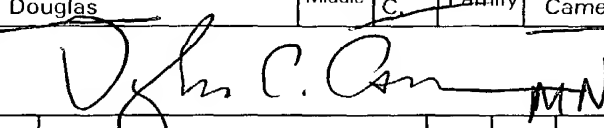
Please type a plus sign (+) inside this box ☐

DECLARATION				Page 2	
<p>I hereby claim benefit under Title 35, United States Code § 120 of any United States application(s), or § 365(C) of any PCT international application designating the United States of America, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application or PCT international application in the manner provided in the first paragraph of Title 35, United States Code § 112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations § 1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application.</p>					
U.S. Parent Application Number	PCT Parent Number	Parent Filing Date (MM/DD/YYYY)	Parent Patent Number (if applicable)		
	PCT/US00/23878				
<input type="checkbox"/> Additional U.S. or PCT international application numbers are listed on a supplemental priority sheet attached hereto					
<p>As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and all continuation and divisional applications based thereon, and to transact all business in the Patent and Trademark Office connected therewith.</p>					
<input checked="" type="checkbox"/> Firm Name		<input type="text" value="Quarles & Brady LLP"/>		Customer Number or label <input type="text" value="26734"/>	
<input type="checkbox"/> List attorney(s) and/or agent(s) name and registration number below					
Name	Registration Number	Name	Registration Number		
<input type="checkbox"/> Additional attorney(s) and/or agents named on a supplemental priority sheet attached hereto					
Please direct all correspondence to		<input type="checkbox"/> Customer Number or label		OR <input checked="" type="checkbox"/> Fill in correspondence address below	
Name	Nicholas J. Seay				
Address	Quarles & Brady LLP				
Address	P O Box 2113				
City	Madison		State	WI	
Country	USA		Telephone	(608)251-5000	
			Fax	(608)251-9166	
<p>I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.</p>					
Name of Sole or First Inventor:			A petition has been filed for this unsigned inventor		
Given	Patrick	Middle	F.	Family	Suthers
					Suffix
Inventor's Signature	<i>Patrick F. Suthers</i>				Date
	WI				2002-09-03
Residence:	Madison		State	WI	
			Country	US	
Post Office	806 Olin Ave., Apt. 1				
Post Office					
City	Madison		State	WI	
			Zip	53715	
			Country	US	
			Applicant Authority		
<input checked="" type="checkbox"/> <input checked="" type="checkbox"/> Additional inventors are being named on supplemental sheet(s) attached hereto					

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p99317wo

Please type a plus sign (+) inside this box ☐

2a

DECLARATION										ADDITIONAL INVENTOR(S) Supplemental Sheet											
Name of Additional Joint Inventor, if any:										A petition has been filed for this unsigned inventor											
Given	Douglas				Middle	C.		Family	Cameron				Suffix								
Inventor's											Date	8/26/02									
Residence:	N. Plymouth				State	MN		Country	US				Citizenship	US							
Post Office	3590 Ranier Lane, N.																				
Post Office																					
City	N. Plymouth				State	MN		Zip	55447		Country	US				Applicant Authority					
Name of Additional Joint Inventor, if any:										A petition has been filed for this unsigned inventor											
Given					Middle Initial			Family Name					Suffix								
Inventor's											Date										
Residence:					State			Country					Citizenship								
Post Office																					
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Name of Additional Joint Inventor, if any:										A petition has been filed for this unsigned inventor											
Given					Middle			Family					Suffix								
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Residence:					State			Country					Citizenship								
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Name of Additional Joint Inventor, if any:										A petition has been filed for this unsigned inventor											
Given					Middle			Family					Suffix								
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Residence:					State			Country					Citizenship								
Post Office																					
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Name of Additional Joint Inventor, if any:										A petition has been filed for this unsigned inventor											
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Residence:					State			Country					Citizenship								
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Additional inventors are being named on supplemental sheet(s) attached hereto																					